Accepted Manuscript

Title: Estimation of muscle activation during different walking speeds with two mathematical approaches compared to surface EMG



Authors: Ursula Trinler, Fabien Leboeuf, Kristen Hollands, Richard Jones, Richard Baker

PII:	S0966-6362(18)30285-6
DOI:	https://doi.org/10.1016/j.gaitpost.2018.06.115
Reference:	GAIPOS 6232
To appear in:	Gait & Posture
Received date:	27-3-2018
Revised date:	22-5-2018
Accepted date:	17-6-2018

Please cite this article as: Trinler U, Leboeuf F, Hollands K, Jones R, Baker R, Estimation of muscle activation during different walking speeds with two mathematical approaches compared to surface EMG, *Gait and Posture* (2018), https://doi.org/10.1016/j.gaitpost.2018.06.115

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Estimation of muscle activation during different walking speeds with two

mathematical approaches compared to surface EMG.

Ursula Trinler^{a,b}, Fabien Leboeuf^a, Kristen Hollands^a, Richard Jones^a, Richard Baker^a

^aUniversity of Salford, School of Health Science, Allerton Building, Frederick Road Campus, Salford, M6 6PU, United Kingdom

^bBG Unfallklinik Ludwigshafen, Zentrum für Bewegungsanalytik, Forschung und Lehre, Ludwig-Guttmann Straße 13, 67071 Ludwigshafen, Germany

Corresponding author:

Ursula Trinler

Forschung und Lehre

BG Unfallklinik Ludwigshafen

Ludwig-Guttmann Str. 13

67071 Ludwigshafen

Germany

Ursula.trinler@bgu-ludwigshafen.de, 0049-621 6810 8498

HIGHLIGHTS

- Results show high deviations between modelling approaches and EMG.
- Slower speeds resulted in smaller deviations than faster speeds.
- Neither modelling approach (SO, CMC) had an overall better agreement with EMG.
- Muscle estimation needs further improvements before it can be implemented in CGA.

Abstract

Background: Muscle force estimation could improve clinical gait analysis by enhancing insight into causes of impairments and informing targeted treatments. However, it is not currently standard practice to use muscle force models to augment clinical gait analysis, partly, because robust validations of estimated muscle activations, underpinning force modelling processes, against recorded electromyography (EMG) are lacking. *Research Question:* Therefore, in order to facilitate future clinical use, this study sought to validate estimated lower limb muscle activation <u>using two mathematical models (static optimisation SO, computed muscle control CMC)</u> against recorded muscle activations of ten healthy participants.

Methods: Participants walked at five speeds . Visual agreement in activation onset and offset as well as linear correlation (r) and mean absolute error (MAE) between models and EMG were evaluated.

Results: MAE between measured and recorded activations were variable across speeds (SO vs EMG 15-68%, CMC vs EMG 13-69%). Slower speeds resulted in smaller deviations (mean MAE < 30%) than faster speeds. Correlation was high (r > 0.5) for only 11/40 (CMC) and 6/40 (SO) conditions (muscles X speeds) compared to EMG.

Significance: Modelling approaches do not yet show sufficient consistency of agreement between estimated and recorded muscle activation to support recommending immediate clinical adoption of muscle force modelling. This may be because assumptions underlying muscle activation estimations (e.g. muscles' anatomy and <u>maximum voluntary contraction</u>) are not yet sufficiently individualizable. Future research needs to find timely and cost efficient ways to scale musculoskeletal models for better individualisation to facilitate future clinical implementation.

Keywords: Muscle activation, modelling, surface EMG, walking

Introduction

Neuro-musculoskeletal impairments occurring as a consequence of disease, injury or aging are a substantial burden on our health care system [1]. Clinical gait analysis aids in identifying and understanding the causes of these impairments by measuring the movement and torque of the joints [2]. Knowing the activation and force profiles of individual muscles during various movements can help to gain further insight into the causes of impairments and inform targeted treatments. Yet, a range of muscle activation and force modelling approaches have been applied in research studies of sport performance or clinical interventions [3-7] but use of modelling has not yet become established as a part of routine clinical gait analysis [8].

One reason muscle force modelling has not yet been used to its full advantage in clinical practice is that there is a huge variety of approaches producing different results [8]. The influence of different modelling assumptions and approaches on model outputs are still not fully understood. Each approach incorporates different aspects of musculoskeletal morphology, kinematics and kinetics and muscle function using a range of different assumptions [5, 9, 10]. Estimation techniques range from solving a static optimisation (*SO*) to complex optimal control problems [10]. Other techniques, for example computed muscle control (CMC), combine inverse and forward dynamics [11-13] to track recorded kinematics while improving computational time. Whilst faster than CMC, <u>SO</u> does not incorporate the excitation-contraction dynamics of the muscle [13], which might be crucial for patients with an impaired neuromuscular physiology [14].

Another reason for lack of application of force modelling in clinical gait analyses maybe related to the difficulty in validating estimated muscle forces [15]. This, in turn, inhibits

selection of the approach which may best model the neuro-muscular impairments in the patients it is applied to. Validation of muscle activation is important to underpin valid force outcomes [16] because direct validation of estimated muscle force is only possible with invivo force transducers. Studies have validated force estimations indirectly against EMG [9, 10, 14, 17]. However, this approach is limited by the fact that there is no simple relationship between activation and force [18]. Arguably, a more robust phased approach to validation of force modelling is to first validate its estimated muscle activations against recorded EMG in a range of walking speeds and patient groups. A few studies [9, 19, 20] have made some attempts to validate estimated muscle activation methods against EMG, however, were limited in either how EMG as a validation tool was used (e.g. only visual inspection or using input EMG to validate outputs) or in their modelling approaches).

Additionally, an examination of estimated muscle activations at a range of walking speeds is lacking in the literature. To ensure force modelling is appropriate for clinical use it is particularly important to know whether the estimated activations respond in the same way as experimental EMG at different gait speeds. A prevalent gait impairment of neurological and musculoskeletal conditions is reduced walking speed [21]. Differences in walking speed result in changes in joint angles [22], ground reaction forces (GRF) [23] and EMG [22]. The change in joint kinematics can lead to differences in the contractile state (fibre length, velocity) [24] of each muscle and, therefore, lead to differences in the generation of muscle forces [19]. Therefore, this study seeks to expand the current literature and robustly validate estimated muscle activations underpinning muscle force models by comparing estimated lower limb muscle activation *using SO and CMC* with recorded EMG of ten healthy participants while walking at five speeds . This is a necessary step to validating force modelling and facilitating

its future clinical use. It was hypothesised that muscle activations will generally increase with walking speed, while there will be a good agreement between mathematical models and surface EMG. Furthermore, CMC will result in better agreement with EMG due to the forward dynamic approach compared to <u>SO</u>.

Methods

Ethical approval was granted by the College of Health and Social Care Ethics Panel. Ten healthy adults (5F/5M, mean±SD of age 28±5 years, height 1.72±0.08m, mass 69±12kg) were recruited from amongst University students and staff. All participants provided written informed consent.

Experimental setup

A ten-camera motion capture system (Vicon Nexus 1.8.5, 100Hz) was used with four force plates embedded into the walkway (Kistler, 1000Hz). Before starting the measurement, cameras were ensured to be coexistent with the force plates by using the *Caltester* approach [25] (< 1° force orientation error, < 3mm CoP displacement error). Surface EMG was collected in parallel with a wireless 16 channel Noraxon system using an in-built low pass filter of 500Hz (DTS receiver, 1000Hz). The SENIAM [22] guidelines were adopted and following preparation of the skin, electrodes were placed on the following muscles : rectus femoris, vastus medialis and lateralis, semitendinosus, tibialis anterior, soleus, and gastrocnemius medialis and lateralis.

Reflective markers were placed on following anatomical landmarks similar to a CAST model [26] and adapted from the proposed model gait2392 in OpenSim [27]: acromion, anterior superior iliac spine, midpoint of both posterior superior iliac spines, three markers on the thigh and shank, medial and lateral epicondyle at the knee, medial and lateral malleolus, heel,

midfoot lateral, toe lateral and medial (1st and 5th metatarsal), and tip of the first toe (Figure 1).

Data collection

Following a static standing trial, the participant walked at his/her self-selected speed over the force plates on a ten-meter walkway, while five valid (individual foot fall entirely on a single force plate) gait cycles for each leg were recorded. This procedure was repeated for 20% and 40% slower and faster walking speeds in following order: self-selected, 20% slow, 40% slow, 20% fast, 40% fast. Speed was monitored with timing gates placed five meters apart the start and end of the walkway. Participants were given feedback on their speed to guide them to walk at the prescribed speed to achieving the target speed within 1%.

Data processing

The raw marker trajectories were pre-processed in Vicon by a customised pipeline to calculate several virtual landmarks and joint centres for static scaling [28]. Gait events were defined via automatic force plate detection and visually verified. The GRFs were filtered in MATLAB (2012b, Mathworks, Matick NA), the kinematics in OpenSim (Release 3.3) [12], both with a 6Hz low-pass 2nd order Butterworth filter. EMG signals were offset corrected, rectified, and filtered with a high-pass 2nd order Butterworth filter.

Musculoskeletal model

We used the generic musculoskeletal model gait2392 of OpenSim [29, 30], which describes a male subject with a body height of 1.80m and a body mass of 75.16kg. It consists of 12 rigid-body segments (torso, pelvis, thigh, shank, talus, foot, toes) which are surrounded by 92

musculo-tendon actuators summing up to 72 muscles (Figure 1) while providing in total 21 degree of freedoms (3 DoF of the upper body, 6 DoF of the pelvis, 3 DoF of the hip, 2 DoF-driven knee, 1 DoF of the ankle). A detailed description of the model can be found elsewhere [29, 30].

Estimation of muscle activation

Processed marker trajectories and experimental GRFs were imported into OpenSim 3.3 using tools from *Lee-Son's Toolbox [31]*. The SimTrack pipeline [12] within OpenSim was used for further calculations. After scaling the model [28], dynamic segment poses were tracked using inverse kinematics. Calculated joint angles and experimental GRFs are further used to estimate muscle excitations with two modelling approaches: i) <u>SO</u> and ii) CMC. These estimation techniques have been chosen due to their frequent use in the literature (<u>SO</u>), its novel approach (CMC), and its independence to EMG, which allows an unbiased comparison with EMG. To ensure essential dynamical consistency for CMC between joint kinematics and GRFs, Residual Reduction Algorithm (RRA) was used [10] to adjust mass properties of the model (i.e. centre of mass of the torso, mass of the body segments) using nonphysical compensatory forces and moments called residuals.

<u>SO</u> (inverse dynamics) resolves the net joint moments into individual muscle forces [10] <u>using force-length velocity relation</u> while minimising squared muscle activations as objective function. The same criteria were implemented in CMC, <u>which are the standard setting in</u> <u>OpenSim for both SO and CMC</u>. While <u>SO</u> is a time independent process, CMC loops the process at time step t forward to the next time step t+T. Therefore, the process is timedependent, however, represents only a limited forward dynamics process due to its

7

independency to all other time steps [13]. This allows the model to stay efficient in computational time which is an important factor for routine processing.

Data analysis

All data were normalised to 100% of the gait cycle after which estimated muscle activation profiles were compared qualitatively via visual inspection to the profiles of recorded muscle activations. One representative trial <u>of the dominant leg</u> of each participant and walking speed was chosen by an experienced researcher according to a clean EMG signal <u>to be able to</u> <u>directly compare individual observed activation with the directly related estimated muscle</u> <u>activation</u>. Recorded and estimated muscle activation were normalised to the maximum activation <u>of the self-selected walking trial</u>. <u>Data were</u> further analysed to calculate the mean absolute error (MAE) between <u>SO</u> and CMC, <u>SO</u> and EMG, as well as CMC and EMG of each of the 101 data points (*n*) [32] to analyse the mean deviation between techniques:

$$MAE = \left(\frac{1}{n}\sum_{i=1}^{n}|A1_{i} - A2_{i}|\right) \tag{1}$$

A1_i represents the muscle activation at the ith time step of the gait cycle of one of the three conditions (<u>SO</u>, CMC, EMG), A2_i the muscle activation at the same time step of one of the other conditions [33]. <u>MAE was then multiplied by 100 to represent the percentage of MAE dependent to 100% activity.</u>

Furthermore, the linear correlation of the 101 data points was analysed using the <u>Spearman</u> correlation coefficient r <u>(non-normal distribution)</u> for each participant and walking speed between <u>SO</u> and CMC, <u>SO</u> and EMG, as well as between CMC and EMG was calculated. The classification of Cohen [34] was used (0.1 < |r| < 0.3 weak correlation, 0.3 < |r| < 0.5 moderate correlation, 0.5 < |r| strong correlation). <u>To be able to analyse speed-related</u> <u>changes in more detail, a descriptive trend-analysis of mean muscle activation, (taken</u>

throughout the, normalised, gait cycle, of fast and slow walking speed, relative to mean muscle activation at self-selected speed) was undertaken.

Results

Muscle <u>peak</u> activation is generally increasing with higher walking speeds <u>for estimated and</u> <u>observed muscle activation</u> (Figure 2, <u>muscle activation profiles normalised to a gait cycle</u>). Only tibialis anterior shows visually low speed-dependence for SO and EMG and nearly none with CMC. Vastus lateralis presents similar changes <u>for different approaches</u> in stance but not in swing, where <u>SO</u> shows nearly no activation for all speeds compared to CMC and EMG. Qualitative inspection <u>shows</u> peak values <u>to be</u> comparable between modelling approaches and compared to surface EMG, except for rectus femoris and semitendinosus, especially for the fast walking speeds. <u>Trend analysis (Figure 3) shows average higher</u> <u>activations at faster speeds than slower speeds for both CMC and SO. Mean of slow (SO,</u> <u>CMC) and very slow (SO, EMG) walking for tibialis anterior as well as slow (CMC) for</u> <u>gastrocnemius lateralis, slow (SO, CMC, EMG) for gastrocnemius medialis, and slow (EMG)</u> <u>and very slow (EMG) for semitendinosus are around 0% and have partial standard deviations</u> <u>above zero.</u>

<u>MAE percentage to 100% muscle activity</u> between modelling approaches and EMG were widely spread across walking speeds (SO vs EMG: 15-68%, CMC vs EMG: 13-69%), while only 26 of 40 and 27 of 40 conditions (muscles X speeds) for <u>SO</u> and CMC, respectively, stayed under 30% (Table 1). Smaller MAE are generally shown for slower walking speeds where self-selected (except muscle rectus femoris) slow and very slow stayed under 30% for all muscles. High linear correlation (r > 0.5 [34]) were only found for 11 of 40 (CMC) and 6 of 40 (SO) conditions (muscles X speeds) compared to EMG (Table 2), while shank muscles

resulted in better correlation (r > 0.5, 15 of 40 conditions) than thigh muscles (r > 0.5, 2 of 40 conditions). Rectus femoris shows negative or zero correlation between estimated and recorded muscle activation.

Both, MAE and linear correlation showed better results when comparing modelling approaches than when compared them to EMG.

Discussion

This study aimed to compare estimated muscle activations of <u>SO</u> and CMC modelling approaches to recorded EMG of lower limb muscles in ten healthy participants walking at five different speeds. Such robust validation of estimated muscle activation patterns has been lacking in the literature and is necessary to underpin valid force outcomes and facilitate future use of force modelling to augment clinical gait analyses. Our results show high deviations between modelling approaches and EMG; neither modelling approach had an overall better agreement with EMG. The strongest agreement with EMG, for both modelling approaches, occurred at slower speeds but only for some of the muscles (especially those on the shank). We further considered the response of estimated muscle activations to speed and agreement to EMG of particular muscles to identify how muscle activation estimation can be improved.

As hypothesised, magnitude of both estimated and recorded muscle activations increase with faster walking speeds in all muscles, except for <u>some of</u> tibialis anterior <u>peak muscle</u> <u>activation (Figure 2)</u>. This muscle reacted differently to speed as slower walking resulted in partially higher estimated and recorded activation levels than faster walking, <u>especially for</u> <u>CMC and SO after loading response</u>. Also, slower walking speeds showed similar averaged <u>activation compared to that of self-selected walking speed (Figure 3)</u>. Higher or equivalent

activation of tibialis anterior at slowest walking speeds has also been reported previously [19, 20]. The higher activation for tibialis anterior while walking slower might be caused by a deviation from the natural walking pattern of the participant (i.e. higher energy costs [35]). This might increase or change the demand on the activation of tibialis anterior. Overall, these results show similar changes dependent on walking speed for both estimated and recorded muscle activity.

Slower walking resulted in smaller absolute mean error between estimated and recorded muscle activation for both modelling approaches. This may be because joint moments are sensitive to accelerations [36], which increase with speed and thus amplify the residual between measured and model-calculated GRF. As suggested by Thelen and colleagues [11] we attempted to overcome this problem by introducing a residual reduction algorithm (RRA) before applying CMC which reduces dynamic inconsistency between experimental GRF and kinematic data. While CMC showed stronger correlations for shank muscles, it did not show better overall agreement with recorded EMG compared to \underline{SO} , as might be expected applying RRA before CMC. Therefore, although CMC has the advantage over \underline{SO} to include a limited time-dependent algorithm, CMC cannot be favoured over \underline{SO} according to our results.

Our findings of a general better agreement between modelling approaches and EMG in the shank than thigh muscles is, mostly, in agreement with previous findings of Zuk et al. [17]. This study reported high correlations between modelling approaches and EMG for gastrocnemius or soleus but weaker agreement for biceps femoris, rectus femoris and tibialis anterior. Better agreement between estimated and recorded muscle activation at the shank may be due to better assumptions about individual muscle characteristics in the shank compared to the thigh. Compliance between estimated muscle activations with EMG seems

more dependent on particular characteristics of individual muscles (i.e., lever arm, cross sectional area) rather than estimation technique. Zuk et al. [17], therefore, concluded that further research needs to focus on the force-sharing problem (which muscle is favoured above the other) and the development of cost functions describing motor control strategies (perception of the environment and consequential motor variability) more closely, which means that more effort needs to be undertaken to improve subject-specific musculoskeletal models.

At the level of the thigh, a negative correlation was seen between estimated activation and EMG for rectus femoris. This is due to anticyclic activation pattern between estimated and recorded activation. Previous literature has shown, that rectus femoris might be only active around transition from stance to swing, with recorded activations at the start and end of a gait cycle being attributed to crosstalk from the vastii muscles [37]. This indicates a potential limitation of using surface EMG as a validation tool for estimating muscle activation; as crosstalk from other muscles may confound the validation process [32]. Further, this highlights the importance of good quality EMG collections in order to confirm estimated activations.

Summarised, some other limitations of the present study are related to the limited ability to adjust the musculoskeletal model. Only segment length, mass and inertia were adjusted to match participants. Muscle specific characteristics, like the individual maximum muscle force-length relationships of muscle and tendon, and their origins and insertions have been kept according to the generic model. This may have made for less accurate muscle activation estimations. Furthermore, <u>SO</u> is based on a rigid tendon assumption, while CMC is not. Again, this might have been expected to induce differences between models [38]. However,

12

neither model agreed with recorded EMG more closely or consistently than the other so this is not thought to be a major limitation for a healthy population [10]. Finally, when considering clinical use of modelling approaches, use of optimisation criterion (minimisation of the squared sum of all muscle activation), which do not take co-contraction into consideration, may limit validity in application to patients with higher activation of agonistic and antagonistic muscles [39].

Conclusion

The presented difference between estimated and recorded muscle activation and non-linear correlation for some of the muscles indicates that estimation of muscle activations, and, therefore, estimation of muscle forces, need further improvements before they can be implemented in a routine clinical setting. Neither <u>SO</u> nor CMC resulted in better agreement to EMG, and can be favoured above the other. More research on patients is needed to understand relevant advantages of different mathematical estimation techniques. For this, future research needs to find timely and cost efficient ways to scale musculoskeletal models for a better individualisation to facilitate future clinical implementation. Specifically, further development in subject specific muscle modelling and the force-sharing problem are crucial that estimation errors can be minimised.

Conflict of interest statement

All authors had no financial and personal relationships with other people or organisations that could inappropriately influence (bias) this work.

References

[1] Woolf AD, Erwin J, March L. The need to address the burden of musculoskeletal conditions. Best Pract Res Clin Rheumatol. 2012;26:183-224.

[2] Davis RB. Reflections on clinical gait analysis. Journal of electromyography and kinesiology : official journal of the International Society of Electrophysiological Kinesiology. 1997;7:251-7.

[3] Buchanan TS, Lloyd DG, Manal K, Besier TF. Estimation of Muscle Forces and Joint Moments Using a Forward-Inverse Dynamics Model. Medicine & Science in Sports & Exercise. 2005;37:1911-6.

[4] Arnold AS, Anderson FC, Pandy MG, Delp SL. Muscular contributions to hip and knee extension during the single limb stance phase of normal gait: a framework for investigating the causes of crouch gait. Journal of biomechanics. 2005;38:2181-9.

[5] Anderson FC, Pandy MG. A dynamic optimization solution for vertical jumping in three dimensions. Comput Meth Biomech Biomed Eng. 1999;2:201-31.

[6] Hatze H. A comprehensive model for human motion sumulation and its application to the take-off phase of the long jump. Journal of biomechanics. 1981;14:135-42.

[7] Alexander N, Schwameder H. Effect of sloped walking on lower limb muscle forces. Gait & posture. 2016;47:62-7.

[8] Trinler U, Hollands K, Jones R, Baker R. A systematic review of approaches to modelling lower limb muscle forces during gait: Applicability to clinical gait analyses. Gait & posture. 2018;61:353-61.

[9] Lin YC, Dorn TW, Schache AG, Pandy MG. Comparison of different methods for estimating muscle forces in human movement. Proceedings of the Institution of Mechanical Engineers Part H - Journal of Engineering in Medicine. 2012;226:103-12.

[10] Anderson FC, Pandy MG. Static and dynamic optimization solutions for gait are practically equivalent. Journal of biomechanics. 2001;34:153-61.

[11] Thelen DG, Anderson FC. Using computed muscle control to generate forward dynamic simulations of human walking from experimental data. Journal of biomechanics. 2006;39:1107-15.

[12] Delp SL, Anderson FC, Arnold AS, Loen P, Habib A, John CT, et al. OpenSim: Open-Source Software to Create and Analyze Dynamic Simulations of Movement. IEEE transactions on bio-medical engineering. 2007;54:1940-50.

[13] Thelen DG, Anderson AE, Delp SL. Gererating dynamic simulations of movement using computed muscle control. Journal of biomechanics. 2003;36:321-8.

[14] De Groote F, Pipeleers G, Jonkers I, Demeulenaere B, Patten C, Swevers J, et al. A physiology based inverse dynamic analysis of human gait: potential and perspectives. Computer Methods in Biomechanics & Biomedical Engineering. 2009;12:563-74.

[15] Lund ME, de Zee M, Andersen MS, Rasmussen J. On validation of multibody musculoskeletal models. Proceedings of the Institution of Mechanical Engineers Part H, Journal of engineering in medicine. 2012;226:82-94.

[16] Hicks J, Uchida TK, Seth A, Rajagopal A, Delp SL. Is my model good enough? Best practices for verification and validation of musculoskeletal models and simulations of movement. Journal of biomechanical engineering. 2015;137:020905.

[17] Żuk M, Syczewska M, Pezowicz C. Use of the surface electromyography for a quantitative trend validation of estimated muscle forces. Biocybernetics and Biomedical Engineering. 2018.

[18] Erdemir A, McLean S, Herzog W, van den Bogert AJ. Model-based estimation of muscle forces exerted during movements. Clinical Biomechanics. 2007;22:131-54.

[19] Neptune RR, Sasaki K, Kautz SA. The effect of walking speed on muscle function and mechanical erergetics. Gait & posture. 2008;28:135-43.

[20] Liu MQ, Anderson FC, Schwartz MH, Delp SL. Muscle contributions to support and progression over a range of walking speeds. Journal of biomechanics. 2008;41:3243-52.

[21] Dickstein R. Rehabilitation of gait speed after stroke: a critical review of intervention approaches. Neurorehabilitation and neural repair. 2008;22:649-60.

[22] Schwartz MH, Rozumalski A, Trost JP. The effect of walking speed on the gait of typically developing children. Journal of biomechanics. 2008;41:1639-50.

[23] Lelas JL, Merriman GJ, Riley PO, Kerrigan DC. Predicting peak kinematic and kinetic parameters from gait speed. Gait & posture. 2003;17:106-12.

[24] Arnold EM, Hamner SR, Seth A, Millard M, Delp SL. How muscle fiber lengths and velocities affect muscle force generation as humans walk and run at different speeds. J Exp Biol. 2013;216:2150-60.

[25] Holden J, Selbie W, Stanhope S. A proposed test to support the clinical movement analysis laboratory accreditation process. Gait & posture. 2003;17:205-13.

[26] Leardini A, Sawacha Z, Paolini G, Ingrosso S, Nativo R, Benedetti MG. A new anatomically based protocol for gait analysis in children. Gait & posture. 2007;26:560-71.

[27] Chand J, Hammer S, Hicks J. Collecting Experimental Data. OpenSim Documentation2012.

[28] Trinler U, Baker R. Estimated landmark calibration of biomechanical models for inverse kinematics. Medical engineering & physics. 2017.

[29] Delp SL. Surgery Simulation: A Computer Graphics System to Analyse and Design Musculoskeletal Reconstructions of the Lower Limb [Ph.D.]. Stanford, CA: Standford University; 1990.

[30] Delp SL, Loan P, Hoy MG, Zajac FE, Topp EL, Rosen JM. An interactive graphicsbased model of the lower extremtly to study orhtopaedics surgical procedures. IEEE transactions on bio-medical engineering. 1990;37:757-67.

[31] Lee S, Son J. Lee-Son's Toolbox: a Toolbox that Converts VICON Mocap Data into OpenSim Inputs. 2010.

[32] Alexander N, Schwameder H. Comparison of Estimated and Measured Muscle Activity During Inclined Walking. Journal of applied biomechanics. 2016;32:150-9.

[33] de Zee M, Dalstra M, Cattaneo PM, Rasmussen J, Svensson P, Melsen B. Validation of a musculo-skeletal model of the mandible and its application to mandibular distraction osteogenesis. Journal of biomechanics. 2007;40:1192-201.

[34] Cohen J. A Power Primer. Psychological Bulletin. 1992;112:155-9.

[35] Waters R, Lunsford B, Perry J, Byrd R. Energy - Speed Relationship of Walking: Standard Tables. Journal of Orthopaedic Research. 1988;6:215-22.

[36] Cahouet V, Luc M, David A. Static optimal estimation of joint acceleration for inverse dynamics problem solution. Journal of biomechanics. 2002;35:1507-13.

[37] Nene A, Byrne C, Hermens H. Is rectus femoris really a part of quadriceps?: Assessment of rectus femoris function during gait in able-bodied adults. Gait & amp; Posture. 2004;20:1-13.

[38] De Groote F, Kinney AL, Rao AV, Fregly BJ. Evaluation of Direct Collocation Optimal Control Problem Formulations for Solving the Muscle Redundancy Problem. Annals of biomedical engineering. 2016;44:2922-36.

[39] Heiden TL, Lloyd DG, Ackland TR. Knee joint kinematics, kinetics and muscle cocontraction in knee osteoarthritis patient gait. Clinical Biomechanics. 2009;24:833-41.

Figure legends

FIGURE 1. Musculoskeletal model gait2392 with applied marker model.



FIGURE 2a. Mean estimated muscle activation <u>of the shank</u> of 10 participants using static optimisation <u>(SO)</u> and <u>computed muscle control</u> (CMC) compared to surface EMG for all five walking speeds. Speeds are indicated from light grey (very slow) to black (very fast). Time (x-axis) is normalised to 100% of a gait cycle, muscle activation (y-axis) is normalised to peak activation of a typical self-selected walking trial.

FIGURE 2b. Mean estimated muscle activation <u>of the thigh</u> of 10 participants using static optimisation <u>(SO)</u> and <u>computed muscle control</u> (CMC) compared to surface EMG for all five walking speeds. Speeds are indicated from light grey (very slow) to black (very fast). Time (x-axis) is normalised to 100% of a gait cycle, muscle activation (y-axis) is normalised to peak activation of a typical self-selected walking trial.





FIGURE 3. Trend-analysis of mean muscle activation averaged over ten participants, taken throughout the normalised gait cycle, of fast and slow walking speed, relative to mean muscle activation at self-selected speed in percentage.



TABLE 1. Mean absolute error and its SD	(activation level in %)) between static o	ptimisation
---	-------------------------	--------------------	-------------

Speed	Method	ТА	GM	GL	Sol	VM	VL	RF	ST
<u> </u>			-	-					
VF	SO-EMG	41 (8)	33 (7)	42 (13)	46 (14)	71 (29)	65 (25)	68 (29)	49 (20)
	CMC-EMG	32 (8)	31 (7)	40 (12)	38 (14)	57 (27)	55 (24)	69 (27)	49 (18)
	SO-CMC	22 (3)	12 (3)	19 (7)	10 (2)	34 (14)	34 (15)	37 (16)	43 (21)
F	SO-EMG	29 (7)	27 (7)	28 (8)	33 (11)	41 (14)	38 (8)	46 (22)	31 (18)
	CMC-EMG	21 (4)	26 (11)	28 (8)	28 (11)	34 (13)	32 (9)	50 (22)	32 (15)
	SO-CMC	22 (1)	13 (2)	17 (5)	8 (2)	25 (7)	24 (8)	30 (12)	30 (17)
Ν	SO-EMG	24 (6)	25 (6)	19 (4)	27 (6)	27 (7)	23 (4)	35 (7)	21 (6)
	CMC-EMG	19 (4)	23 (9)	20 (5)	23 (7)	22 (5)	20 (5)	34 (6)	22 (7)
	SO-CMC	20 (1)	12 (2)	13 (3)	7 (2)	22 (6)	20 (6)	27 (6)	21 (7)
S	SO-EMG	23 (8)	21 (4)	15 (5)	22 (8)	18 (9)	17 (8)	23 (9)	18 (6)
	CMC-EMG	17 (4)	21 (7)	20 (6)	20 (7)	16 (6)	15 (10)	24 (9)	19 (7)
	SO-CMC	21 (6)	10 (2)	13 (2)	5 (1)	18 (7)	16 (8)	18 (9)	16 (7)
VS	SO-EMG	24 (15)	20 (6)	15 (5)	19 (8)	15 (8)	15 (7)	21 (9)	19 (7)
	CMC-EMG	19 (10)	20 (9)	18 (4)	17 (7)	13 (5)	13 (7)	21 (11)	19 (6)
	SO-CMC	18 (6)	10 (2)	13 (3)	5 (1)	16 (7)	15 (7)	18 (9)	17 (8)

(SO) and computed muscle control (CMC), SO and EMG, and CMC and EMG.

VF= very fast, F= fast, N= self-selected, S= slow, VS= very slow; SO= static optimisation, CMC= computed muscle control, TA = tibialis anterior, $\underline{GM} = \underline{gastrocnemius medialis}$, GL= gastrocnemius lateralis, $\underline{Sol} = \underline{soleus}$, $\underline{VM} = \underline{vastus medialis}$, VL= vastus lateralis, RF= rectus femoris, ST = semitendinosus.

Speed	Method	TA	GM	GL	<u>Sol</u>	VM	VL	RF	ST
		0.31	0.45	0.53	0.33	0.10	0.14	-0.15	0.38
VF	SO-EMG	(0.28)	(0.27)	(0.17)	(0.24)	(0.14)	(0.23)	(0.23)	(0.37)
	CMC-	0.61	0.55	0.62	0.59	0.26	0.21	-0.25	0.14
	EMG	(0.26)	(0.34)	(0.22)	(0.28)	(0.30)	(0.35)	(0.14)	(0.32)
		0.51	0.93	0.95	0.88	0.60	0.61	0.46	0.39
	SO-CMC	(0.10)	(0.03)	(0.03)	(0.06)	(0.26)	(0.26)	(0.32)	(0.32)
		0.37	0.33	0.46	0.19	0.07	0.15	-0.05	0.48
F	SO-EMG	(0.23)	(0.28)	(0.19)	(0.22)	(0.15)	(0.23)	(0.25)	(0.30)
	CMC-	0.62	0.49	0.52	0.35	0.15	0.22	-0.17	0.18
	EMG	(0.18)	(0.34)	(0.27)	(0.29)	(0.31)	(0.31)	(0.29)	(0.30)
		0.50	0.90	0.90	0.92	0.52	0.53	0.54	0.41
	SO-CMC	(0.08)	(0.05)	(0.03)	(0.04)	(0.20)	(0.20)	(0.25)	(0.38)
		0.40	0.26	0.52	0.17	0.17	0.33	-0.13	0.52
Ν	SO-EMG	(0.26)	(0.28)	(0.19)	(0.17)	(0.20)	(0.27)	(0.23)	(0.22)
	CMC-	0.60	0.47	0.46	0.31	0.22	0.37	-0.20	0.21
	EMG	(0.15)	(0.33)	(0.22)	(0.22)	(0.27)	(0.35)	(0.28)	(0.27)
		0.48	0.87	0.84	0.92	0.45	0.46	0.33	0.49
	SO-CMC	(0.10)	(0.07)	(0.08)	(0.03)	(0.29)	(0.27)	(0.28)	(0.33)
		0.32	0.38	0.63	0.25	0.17	0.33	0.04	0 54
S	SO-FMG	(0.32)	(0.24)	(0.11)	(0.23)	(0.18)	(0.28)	(0.19)	(0.21)
D	CMC-	0.66	0.52	0.45	0.30	0.23	0.34	-0.12	0.19
	EMG	(0.16)	(0.32)	(0.10)	(0.23)	(0.37)	(0.38)	(0.12)	(0.26)
	Line	0.43	0.89	0.84	0.95	0.44	0.43	0.33	0.41
	SO-CMC	(0.14)	(0.04)	(0.06)	(0.03)	(0.339	(0.32)	(0.35)	(0.27)
		0.22	0.41	0.60	0.20	0.19	0.28	0.06	0.45
VS	SO EMG	(0.33)	(0.41)	(0.10)	(0.15)	(0.18)	(0.28)	-0.00	(0.45)
v S	CMC	(0.33)	(0.27)	(0.10)	(0.13)	(0.23)	(0.20)	(0.20)	(0.20)
	EMG	(0.13)	(0.31)	(0.34)	(0.10)	(0.37)	(0.31)	-0.10	(0.25)
	LINIO	0.15)	0.87	(0.34)	(0.15)	(0.37)	(0.30)	(0.17)	(0.23)
	SO-CMC	(0.11)	(0.07)	(0.12)	(0.03)	(0.35)	(0.34)	(0.36)	(0.33)
	EMG SO-CMC	(0.13) 0.49 (0.11)	(0.30) 0.87 (0.07)	(0.34) 0.77 (0.12)	(0.19) 0.96 (0.03)	(0.37) 0.44 (0.35)	(0.30) 0.41 (0.34)	(0.19) 0.32 (0.36)	(0.25) 0.37 (0.33)

TABLE 2. Mean correlation coefficient and SD between muscle activation of static

optimisation (SO) and computed muscle control (CMC), SO and EMG, and CMC and EMG.

VF= very fast, F= fast, N= self-selected, S= slow, VS= very slow; SO= static optimisation, CMC= computed muscle control, TA = tibialis anterior, $\underline{GM = gastrocnemius medialis}$, GL= gastrocnemius lateralis, $\underline{Sol = soleus}$, $\underline{VM = vastus medialis}$, VL= vastus lateralis, RF= rectus femoris, ST = semitendinosus.