

1 ORIGINAL ARTICLE

2 **Capuchin monkey biogeography: understanding *Sapajus* Pleistocene range**  
3 **expansion and the current sympatry between *Cebus* and *Sapajus***

4

5 Marcela G. M. Lima<sup>1,2</sup>, Janet C. Buckner<sup>3</sup>, José de Sousa e Silva-Júnior<sup>2</sup>, Alexandre  
6 Aleixo<sup>2</sup>, Amely Martins<sup>4,5</sup>, Jean P. Boubli<sup>6</sup>, Andrés Link<sup>7</sup>, Izeni P. Farias<sup>8</sup>, Maria  
7 Nazareth da Silva<sup>9</sup>, Fabio Röhe<sup>9,10</sup>, Helder Queiroz<sup>11</sup>, Kenneth L. Chiou<sup>12</sup>, Anthony Di  
8 Fiore<sup>4</sup>, Michael E. Alfaro<sup>1,3</sup>, Jessica W. Lynch Alfaro<sup>1,13</sup>

9

- 10 1. Institute for Society and Genetics, University of California, Los Angeles, CA,  
11 USA
- 12 2. Curso de Pós-Graduação em Zoologia, Universidade Federal do Pará/Museu  
13 Paraense Emílio Goeldi, Belém, PA, Brazil
- 14 3. Department of Ecology and Evolutionary Biology, University of California, Los  
15 Angeles, USA
- 16 4. Department of Anthropology, University of Texas at Austin, Austin, TX, USA
- 17 5. Centro Nacional de Pesquisa e Conservação de Primatas Brasileiros, ICMBio,  
18 MMA, Brazil
- 19 6. School of Environment and Life Sciences, University of Salford, UK
- 20 7. Department of Biological Sciences and School of Management, Universidad de  
21 Los Andes, Bogotá, Colombia
- 22 8. Universidade Federal do Amazonas UFAM, Manaus, AM, Brazil
- 23 9. Instituto Nacional de Pesquisas da Amazonia (INPA), Manaus, AM, Brazil
- 24 10. Wildlife Conservation Society, Manaus, Brazil
- 25 11. Instituto de Desenvolvimento Sustentável Mamirauá, Tefé, AM, Brazil

26 12. Washington University in St. Louis, St. Louis, MO, USA

27 13. Department of Anthropology, UCLA, Los Angeles, CA, USA

28

29 Correspondence: Marcela G. M. Lima

30 Museu Paraense Emílio Goeldi. Departamento de Mastozoologia. Av. Perimetral, 1901

31 - Terra Firme. Belém, PA, Brasil, CEP 66040-170

32

33 E-mail: marcela\_gml@yahoo.com.br

34

35 SHORT RUNNING HEAD: [max. 8 words] Capuchin monkey biogeography

36

37 WORD COUNT: 6304 (abstract, main text, references)

38

39 **ABSTRACT**

40

41 **Aim**

42 Our aim was to examine gracile capuchin (*Cebus*) and robust capuchin monkey  
43 (*Sapajus*) diversification, with a focus on recent *Sapajus* expansion within Amazonia.

44 We wanted to reconstruct the biogeographical history of the clade using statistical  
45 methods that model lineages' occupation of different regions over time, to evaluate  
46 recently proposed "Out of Amazonia" and "Reinvasion of Amazonia" hypotheses as  
47 alternative explanations for the extensive geographic overlap between reciprocally  
48 monophyletic gracile (*Cebus*) and robust (*Sapajus*) capuchin monkeys.

49

50 **Location**

51 Central and South America.

52

### 53 **Methods**

54 We reconstructed a time-calibrated molecular phylogeny for capuchins under Bayesian  
55 inference from three mitochondrial genes. We then categorized 12 capuchin clades  
56 across four Neotropical centres of endemism and reconstructed the biogeographic  
57 history of the capuchin radiation using six models implemented in ‘BioGeoBEARS’.  
58 We performed a phylogeographic analysis for a robust capuchin clade that spans the  
59 Atlantic Forest, Cerrado, Caatinga and Amazonia.

60

### 61 **Results**

62 We find support for a late Miocene vicariant *Cebus-Sapajus* divergence and a  
63 Pleistocene *Sapajus* invasion of Amazonia from the Atlantic Forest. Our new analyses  
64 confirm *Sapajus* diversified first in the Atlantic Forest, with subsequent range  
65 expansion into widespread sympatry with *Cebus* in Amazonia, as well as multiple  
66 expansions into drier savanna-like habitats. We do not find mitochondrial molecular  
67 congruence with morphological species distinctions for *Sapajus flavius*, *S. cay*, *S.*  
68 *macrocephalus*, *S. libidinosus* and *S. apella*; instead, these five morphological types  
69 together form a single widespread clade (Bayesian posterior probability = 1) with  
70 geographic substructure and shared ancestry during the Pleistocene.

71

### 72 **Main conclusions**

73 Our results support vicariance dividing ancestral capuchin populations in Amazonia  
74 versus the Atlantic Forest, and a Pleistocene “Amazonian invasion” by *Sapajus* to  
75 explain the present day sympatry of *Cebus* and *Sapajus*.

76

77 **Keywords** Amazonia, Atlantic Forest, BEAST, phylogeography, Caatinga, *Cebus*  
78 *kaapori*, Cerrado, RASP Biogeography, range expansion, *Sapajus flavius*

79

## 80 **INTRODUCTION**

81

82 Neotropical primates are the major diurnal mammalian radiation in the  
83 Neotropics, and include over 140 species found across Amazonia, the Andes, the  
84 Atlantic Forest, and even habitats such as the Llanos, Caatinga, Cerrado and Central  
85 Grasslands that are too hot and dry to support many types of mammals. Neotropical  
86 primate diversity is highest within the tropical rainforest habitat in Amazonia. Major  
87 questions include whether Neotropical primates diversified first in Amazonia, and how  
88 and when they spread throughout the various biomes they now inhabit.

89 The wide distribution of capuchin monkeys, with species ranging from Central  
90 America to northern Argentina, makes them a potential model system for addressing  
91 questions of Neotropical primate biogeography. Capuchins comprise two major forms,  
92 the more lightly built gracile capuchin, *Cebus*, and the robust capuchin, *Sapajus*.  
93 Although recent studies have confirmed the genetic distinctiveness of these two lineages  
94 (Lynch Alfaro *et al.*, 2012a; Martins Jr. *et al.*, 2014), key questions remain regarding  
95 their present day distribution, including the extensive sympatry between Amazonian  
96 *Cebus* and *Sapajus* species.

97 Two competing hypotheses have been posited to explain current capuchin  
98 distributions (Lynch Alfaro *et al.*, 2012a). The “Out of Amazonia” (OOA) hypothesis  
99 places the ancestral range for all capuchins in Amazonia, an initial diversification of  
100 both crown robust and crown gracile capuchins within Amazonia, and subsequent

101 independent and recent invasions of non-Amazonian regions by both gracile and robust  
102 capuchins (Lynch Alfaro *et al.*, 2012a). The “Reinvasion of Amazonia” (ROA)  
103 hypothesis proposes that gracile capuchins, *Cebus*, diverged from robust capuchins,  
104 *Sapajus* early in capuchin evolutionary history; crown *Cebus* originated in Amazonia  
105 and crown *Sapajus* in the Atlantic Forest; and the presence of *Sapajus* in the Amazonia  
106 today reflects a recent invasion from the Atlantic Forest into *Cebus*-occupied territory  
107 (Lynch Alfaro *et al.*, 2012a).

108         Phylogeographic analyses using pure dispersal models strongly support the  
109 “Reinvasion of Amazonia” hypothesis (Lynch Alfaro *et al.*, 2012a), and as such have  
110 transformed the interpretation of capuchin taxonomy, conservation biology,  
111 morphology and ecology (see Lynch Alfaro *et al.*, 2012b, 2014, 2015b; Matthews,  
112 2012; Rylands *et al.*, 2013; Martins Jr. *et al.*, 2014; Oliveira *et al.*, 2014; Boubli *et al.*,  
113 2015; Schneider & Sampaio, 2015; Wright *et al.*, 2015). However, limitations of these  
114 studies have left key predictions of the OOA hypothesis untested. For example, Lynch  
115 Alfaro *et al.* (2012a) and Nascimento *et al.* (2015) provided limited sampling of eastern  
116 Amazonia and Cerrado/Caatinga localities, which have the potential to strongly  
117 corroborate the OOA scenarios. In addition, capuchin monkey biogeography has yet to  
118 be interpreted in light of models that integrate processes beyond pure dispersal  
119 (Ronquist, 1997; Ree *et al.*, 2005; Ree & Smith, 2008; Landis *et al.*, 2013).

120         Here we assemble an expanded sampling of capuchin monkeys across their  
121 range and conduct new biogeographic analyses using recently developed statistical  
122 approaches. Our overall goal is to test hypotheses explaining capuchin biogeography  
123 (Boubli *et al.*, 2012; Lynch Alfaro *et al.*, 2012a; Nascimento *et al.*, 2015; Ruiz-Garcia *et*  
124 *al.*, 2016). We thus assemble a new cytochrome *b* + D-loop + cytochrome oxidase I  
125 matrix with three times the samples and more comprehensive spatial and taxonomic

126 sampling compared to Lynch Alfaro *et al.* (2012a). We produce a detailed phylogeny  
127 for capuchin monkeys, with all major lineages represented, to confirm the monophyly  
128 of *Cebus* and *Sapajus* and to recover major clades within the capuchin radiation. We  
129 compare alternative biogeographical scenarios using a flexible framework for inferring  
130 spatial histories that allows for a number of different mechanisms causing range  
131 expansion or contraction. We assess whether the employment of a more complete  
132 genetic dataset and statistical methods with expanded models increase support for  
133 previous conclusions about capuchin biogeographical evolution or support new  
134 scenarios altogether. Our sampling allows us to address two new biogeographic  
135 hypotheses for this clade. The first hypothesis postulates vicariance between Amazonia  
136 and the Atlantic Forest. Divergence between *Sapajus* and *Cebus* is caused by isolation  
137 of ancestral capuchin populations in the Atlantic Forest versus Amazonian habitats,  
138 respectively, with subsequent expansion of *Sapajus* into sympatry with Amazonian  
139 *Cebus* (“Reinvasion of Amazonia” hypothesis). The second, opposite, hypothesis  
140 postulates vicariance within Amazonia due to the formation of the Amazon River.  
141 Capuchins arise in Amazonia, and subsequently *Cebus* is isolated north of the Amazon  
142 River, with *Sapajus* isolated south of the Amazon River and expanding into the Atlantic  
143 Forest. Sympatry occurs with subsequent crossing of Amazon River in both directions  
144 (“Out of Amazonia” hypothesis).

145

## 146 **MATERIALS AND METHODS**

147

148 *DNA extraction, amplification, sequencing and alignment*

149

150 We sequenced three mitochondrial genes, cytochrome *b* (*Cyt b*, 1133 bp), D-loop  
151 (1244 bp) and cytochrome oxidase I (COI, 673 bp), for 108 capuchin samples from  
152 poorly known or previously unsampled localities and species (see Table S1 in Appendix  
153 S1). For *Cyt b* and D-loop we used primers and PCR protocols from Lynch Alfaro *et al.*  
154 (2012a) and Boubli *et al.* (2012). We adapted COI primers and protocols from Ward *et*  
155 *al.* (2005) (see Appendix S2). We combined the 108 new samples with 40 samples  
156 analysed previously by Lynch Alfaro *et al.* (2012a) and 13 new samples with reliable  
157 species identification extracted from GenBank, totaling 161 capuchin samples. Note  
158 that samples from Ruiz-Garcia *et al.* (2016) were not available for use in this study. We  
159 used only sequences over 500 bp long to improve clade support (Nascimento *et al.*,  
160 2015). We used the most recent comprehensive taxonomic review for capuchins  
161 (Rylands *et al.*, 2013), in relation to specimen morphology and collection locality, to  
162 assign species names to samples (locality data in Figure 1 and Table S1 in Appendix  
163 S1). Although the limitations of mitochondrial DNA studies are well understood, *Cyt b*,  
164 D-loop and COI have performed well in capturing phylogeny and biogeographical  
165 history in a wide range of species (Tobe *et al.*, 2010; Zhang *et al.*, 2011; Robins *et al.*,  
166 2014). In addition, mtDNA sequence can be reliably generated from museum “crusties”  
167 and skins using now standard protocols (e.g., Lynch Alfaro *et al.*, 2012a, 2015a; Mercês  
168 *et al.*, 2015).

169

#### 170 *Phylogenetic Reconstruction*

171

172 We used MRBAYES 3.2.6 (Ronquist *et al.*, 2012) to reconstruct the phylogenetic  
173 relationships among capuchins, using *Saimiri* as an outgroup. We partitioned *Cyt b* and  
174 COI by codon for the analysis and ran the Markov Chain Monte Carlo (MCMC) for 50

175 million generations. We used ‘sumt’ to calculate the Bayesian consensus tree. We then  
176 used BEAST 1.8.1 (Drummond *et al.*, 2012) to reconstruct a time tree appropriate for  
177 our biogeographical analyses by pruning the alignment to a single exemplar taxon for  
178 each recovered well-supported clade. We considered branch lengths and recovered  
179 polytomies in addition to posterior probabilities at nodes when distinguishing clades.  
180 This resulted in a new, pruned alignment with 13 tips, representing 12 capuchin clades  
181 and one outgroup. We specified a HKY + G model of nucleotide substitution and  
182 partitioned by codon, using *Cyt b* only to avoid branch length bias due to missing data  
183 in the other genes. We used *Saimiri* as an outgroup and the fossil *Neosaimiri* (Kay,  
184 2015) to calibrate the split at 12.5 Ma (lognormal prior, offset=12.5, mean=0, SD=1)  
185 between capuchins and squirrel monkeys for the time tree. We ran the MCMC analysis  
186 for 100 million generations, sampling states every 10,000 generations with a Yule prior  
187 on the diversification rate, and an uncorrelated, lognormal relaxed molecular clock.

188

### 189 *Biogeographical modelling*

190

191 *Biogeographical scenarios* In addition to testing for evidence for “Out of Amazonia”  
192 versus “Reinvasion of Amazonia” hypotheses (see Introduction), we tested for  
193 reciprocal monophyly for *Sapajus* in Amazonia (*S. macrocephalus* + *S. apella*); the  
194 Cerrado + Caatinga + Central Grasslands (*S. cay* + *S. libidinosus*); and the Atlantic  
195 Forest (*S. flavius* + *S. xanthosternos* + *S. robustus* + *S. nigritus*). Lack of reciprocal  
196 monophyly across habitats would suggest multiple independent invasions and  
197 convergent adaptations to habitat types by independent *Sapajus* lineages.

198



199 *Historical biogeography* To test alternative hypotheses of capuchin origins and  
200 radiation we used the R-package ‘BioGeoBEARS’, a recently developed flexible  
201 maximum likelihood framework for inferring spatial histories (Matzke, 2013, 2014; see  
202 Buckner *et al.*, 2015 and Lynch Alfaro *et al.*, 2015a for its recent application to primate  
203 taxa). ‘BioGeoBEARS’ software allows for a statistical comparison of a suite of models  
204 that include alternative biogeographic processes related to cladogenesis and that allow  
205 for species distributions across multiple regions (Matzke, 2014).

206 Our tree was pruned to include one representative tip for each ‘species’  
207 recovered in the MRBAYES analysis. We lumped *Sapajus flavius*, *S. cay*, *S. libidinosus*,  
208 *S. apella*, and *S. macrocephalus* (*Sapajus* clade 4, see below) as one species. This  
209 resulted in a new, pruned alignment with 12 tips. We coded presence/absence in four  
210 major geographic regions for each tip based on known species distribution using  
211 Rylands *et al.* (2013): Atlantic Forest (AF), Cerrado/Caatinga/Central Grasslands (CC),  
212 Amazonia (AM), and Central America/Andes (CA). We used the pruned, time-  
213 calibrated tree with regional coding in ‘BioGeoBEARS’ (Matzke, 2013; Matzke, 2014).  
214 Our candidate pool consisted of six models: DEC, DEC-J, DIVA, DIVA-J,  
215 BAYAREA-LIKE, BAYAREALIKE-J and we used model weights calculated from  
216 corrected Akaike information criterion (AICc) to identify the best model or set of  
217 models for our data (Matzke, 2014). ‘BioGeoBEARS’ script is available in Appendix  
218 S2.

219  
220 *Phylogeographical reconstruction of widespread Sapajus* To examine phylogeographic  
221 patterns for a widespread robust capuchin group recovered in the MRBAYES tree, we  
222 performed both symmetric and asymmetric discrete states phylogeographic analyses in  
223 BEAST under a constant size coalescent prior. These analyses allowed us to take into

224 consideration the uncertainty in the relationships by simultaneously estimating the  
225 topology and the distribution history. To compare the asymmetric and symmetric  
226 analyses for their fit to our data, we performed marginal likelihood estimation (MLE)  
227 using path sampling (PS) and stepping stone (SS) sampling (Beale, 2012, 2013). We  
228 used the default priors for the MLE. We categorized the locality of each tip for the  
229 widespread *Sapajus* clade in the Bayesian consensus tree as occurring in one  
230 biogeographical ‘region’ (Figure 2). We divided Amazonia into four regions using the  
231 Amazon River and two major tributaries, Negro and Madeira, as described by Wallace  
232 (1852) and later Cracraft (1985): Guianas (GU), Negro (NE), Inambari (IN), R ndonia  
233 (RO). We also separated the drier open ecosystems Central Grasslands, Cerrado and  
234 Caatinga (CC), from the Atlantic Forest (AF). These regions were modified from  
235 Jameson Kiesling *et al.* (2015) and Cracraft (1985). As mtDNA is inherited maternally,  
236 it tracks the movement of female lineages over time. Capuchin monkeys are female  
237 philopatric with male-biased dispersal (Lynch Alfaro *et al.*, 2014), so tracing female  
238 lineage was a conservative test for the amount of movement over time in this taxon.

239         Within Amazonia, we tested for isolation by river barriers for widespread  
240 *Sapajus*. If gene flow within Amazonian capuchin populations was restricted to major  
241 interfluves, we expected a pattern of closely related subclades within, not across, four  
242 major Amazonian regions delineated by the Amazon River and its major tributaries.  
243 Evidence for the river barrier hypothesis was found for the sister group to capuchins, the  
244 Amazonian squirrel monkeys (Lynch Alfaro *et al.*, 2015a), as well as for marmosets and  
245 tamarins (Buckner *et al.*, 2015). Robust capuchin lineages with clades spanning  
246 multiple interfluves would provide evidence against rivers as significant isolating  
247 mechanisms.

248

249 **RESULTS**

250

251 *Capuchin Phylogenetics*

252

253           We find strong support for reciprocally monophyletic clades of gracile and  
254 robust capuchins in the phylogenetic reconstruction from MRBAYES (Figure 3a). Our  
255 truncated species time tree from BEAST (Figure S1 in Appendix S3) estimates the  
256 mean divergence time between *Sapajus* and *Cebus* at 5.8 Ma [95% higher posterior  
257 density (HPD) = 2.24 - 10.94] (late Miocene).

258           For robust capuchins we recover strong support for the clades *S. xanthosternos*  
259 (posterior probability [pp] = 1) and *S. nigritus* (pp = 1), but little evidence for  
260 reciprocally monophyletic ‘species’ clades for the rest of *Sapajus*. Two *S. robustus*  
261 samples form a weakly supported clade (pp = 0.74). All other robust capuchins form  
262 one large widely distributed clade (‘*Sapajus* CLADE 4,’ pp = 1), comprising *S. apella*,  
263 *S. macrocephalus*, *S. cay*, *S. libidinosus*, and *S. flavius*. Subclades within this group are  
264 not always congruent with morphological hypotheses about robust capuchin species, a  
265 result concordant with the mitochondrial results from Ruiz-García and colleagues  
266 (2012, 2016). Most strikingly we find *S. apella sensu stricto* to be extensively  
267 paraphyletic with respect to other *Sapajus* species. Within the widespread clade, we  
268 recover several geographical subclades: (1) *S. flavius* (pp = 0.95); (2) *S. libidinosus*  
269 together with eastern *S. apella* (pp = 0.80); (3) *S. cay* from Paraguay and Rondônia/Mato  
270 Grosso plus *S. apella* from Guaporé and Apuí (pp = 0.81); (4) *S. cay* from Chapada dos  
271 Guimarães and *S. apella* from Alta Floresta, both in Mato Grosso, Brazil (pp = 1); (5) *S.*  
272 *apella/macrocephalus* (pp = 1) from north of the Amazon and Solimões rivers, as well  
273 as from regions just south of the Amazon River near the Xingu and Tapajos rivers, and

274 just south of Manaus, together reaching all four Amazonian quadrants, as far northwest  
275 as Colombia; (6) *S. apella/macrocephalus* (pp = 0.95) from Brazil and Peru. Minimum  
276 clade ranges for each of the six subclades of the widespread *Sapajus* clade are  
277 geographically coherent though several do not match current morphological hypotheses  
278 about robust capuchin taxonomy (Figure 3c).

279         Within *Cebus*, we recover eight strongly supported clades: *Cebus versicolor*  
280 forms a clade (pp = 1) with *C. cesarae* within the Magdalena river valley in the  
281 Colombian Andes, as part of a larger clade including *C. capucinus* in Panama and  
282 Colombia (pp = 1), and *C. imitator* in Central America (pp = 1). In the eastern Andes,  
283 *C. brunneus* (pp = 1) is sister to *C. leucocephalus* (pp = 1). Within Amazonia, there is  
284 strong support for a *C. unicolor* + *C. albifrons* clade (pp = 1). *C. yuracus* forms a clade  
285 with one *C. unicolor* sample (pp = 1) in western Brazil and Ecuador, and *C. o. olivaceus*  
286 + *C. o. castaneus* + *C. kaapori* form a strongly supported clade (pp = 1). Our *C. kaapori*  
287 sample is from the holotype, collected by Queiroz (1992), from the type locality Chega-  
288 Tudo, in Carutapera Municipality, Maranhão State, Brazil. Here we provide a correction  
289 to the original Queiroz (1992) for the coordinates of that locality, 02°20'S 46°05'W  
290 (Table S1 in Appendix S1).

291

## 292 *Reconstruction of Capuchin Biogeographical History*

293

294         Model weights based on AICc favoured the DEC model (Table 1, Figure 4).  
295 Under this scenario the ancestral capuchin population is inferred to have had a wide  
296 range that included both Amazonia and the Atlantic Forest. The divergence between  
297 *Sapajus* and *Cebus* is a vicariance event, where *Sapajus* is isolated in the Atlantic  
298 Forest, while the lineage leading to gracile *Cebus* is isolated in Amazonia. Crown

299 *Sapajus* is recovered with a distribution restricted to the Atlantic Forest, with  
300 subsequent rapid expansion to the Caatinga/Cerrado/Central Grasslands and Amazonia  
301 regions at about 500 ka. This means that *Cebus*, established in Amazonia at  
302 approximately 2.4 Ma, diversified for about 2 million years before the appearance of  
303 any *Sapajus* clade in that region.

304 For crown *Cebus*, a single expansion event to the Central America/Andes region  
305 occurred at ~2.2 Ma, with subsequent range contractions to just Central America/Andes  
306 for the lineage leading to *C. capucinus*, *C. imitator* and *C. versicolor*, and to Amazonia  
307 only for both the lineage leading to *C. brunneus* (note that the “Amazonia” region  
308 extends to the northern Venezuelan coast; see Figure 4), and for the lineage leading to  
309 *C. unicolor*.

310

### 311 *Reconstruction of widespread Sapajus radiation*

312

313 We do not find any statistical difference in support for the symmetric versus the  
314 asymmetric analyses ( $K \approx 0$ ). Both the phylogeographical reconstructions detail the  
315 explosive Pleistocene range expansion throughout Amazonia, Cerrado, Caatinga and  
316 southern grasslands for *Sapajus* (Figure 2 and Figure S2 in Appendix S3). In the  
317 asymmetric reconstruction, robust capuchins enter Amazonia by a single expansion  
318 from AF to RO at around 300-500 ka, with a second unique expansion across the  
319 Amazon River to GU at 200 ka, and at least two independent expansions across the  
320 Madeira River from RO to IN. There is also evidence for expansion from GU to IN, and  
321 IN to GU, as well as back-crossing from both IN and GU to RO. No strong  
322 geographical pattern demarcates rivers as firm barriers separating the four Amazonian  
323 regions; instead, there has been significant and recurrent recent movement by *Sapajus*

324 across the Amazon, Solimões, Madeira and Negro rivers. *Sapajus* also spread from RO  
325 three times into the Cerrado/Caatinga (CC) region, all within the last 200 ka.

326 The addition of a distribution partition to our BEAST analysis provides support  
327 for the monophyly of *Sapajus robustus* (0.93) and its position as the sister group to a  
328 widespread Amazonian-Grasslands-Atlantic Forest clade (pp = 1.0). *Sapajus robustus* is  
329 restricted to the Atlantic Forest, making it clear that the explosive radiation of robust  
330 capuchin monkeys began with a recent migration into Amazonia and grasslands.

331 A significant difference between our asymmetric and symmetric BEAST  
332 analyses is related to *S. flavius*. In the symmetric BEAST phylogeography analysis, *S.*  
333 *flavius* is recovered as the sister group to *Sapajus* CLADE 4 (Figure S2 in Appendix  
334 S3). On the other hand, *S. flavius* is embedded within the widespread clade in the  
335 asymmetric analysis (Figure 2), suggesting an expansion back into the Atlantic Forest  
336 from the Amazonia/Grasslands regions.

337

## 338 **DISCUSSION**

339

340 Our expanded sample set with comprehensive representation of capuchin species  
341 provides further support for the monophyly of robust and gracile capuchins with a split  
342 at 5.8 Ma. A late Miocene *Sapajus-Cebus* split was previously supported using  
343 combined nuclear and mitochondrial data in Perelman *et al.* (2011) (6.0 Ma, 95%  
344 confidence interval 3.13-9.35 Ma) and Springer *et al.* (2012) (5.7 Ma average, 3.55-8.33  
345 Ma composite 95% min-max). However, these time trees did not include all capuchin  
346 species; here we add *Cebus kaapori*, *C. versicolor*, and *Sapajus flavius*. One exception  
347 to monophyly of *Cebus* and *Sapajus* has been recently reported in Ruiz-Garcia *et al.*  
348 (2016), where some *Cebus albifrons sensu lato* individuals from the Colombian Eastern

349 Llanos and Ecuadorian Amazonia displayed *Sapajus* mitochondrial haplotypes,  
350 suggesting the possibility of localized past or present introgression across these two  
351 genera. Unfortunately, neither sequences nor morphological data are publicly available  
352 for any of these individuals at this time.

353         Our biogeographical analyses reconstruct a widespread capuchin ancestral  
354 distribution across forested areas of South America, from Amazonia to the Atlantic  
355 Forest, supporting Amazonia versus Atlantic Forest vicariant origin for *Cebus* and  
356 *Sapajus*. The distribution of *Sapajus* exclusively in the Atlantic Forest until recently,  
357 and *Cebus* exclusively in the Amazonia and Central America/Andes regions from early  
358 on, suggests a cladogenetic event (vicariance) led to their initial diversification in  
359 isolation.

360         Throughout capuchin evolution, the Brazilian Shield (within AF, CC, RO  
361 Regions) and Guiana Shield (within GU) have been constant highland regions  
362 undergoing few physical changes in configuration but perhaps experiencing changes in  
363 floral assemblages related to climate (Aleixo & Rossetti, 2007). The divergence  
364 between *Sapajus* and *Cebus*, dated consistently at approximately 6 Ma, coincides with  
365 floral assembly of the Cerrado (4-8 Ma), as indicated by the onset of diversification of  
366 many Cerrado plant lineages (Simon *et al.*, 2009). The coincidence in timing of the  
367 divergence between robust and gracile capuchins and of the Cerrado assemblage  
368 suggests that the development of the Cerrado may have acted as a vicariant agent,  
369 especially given the reconstructed distributions of ancestral *Sapajus* (Atlantic Forest)  
370 and *Cebus* (Amazonia). The only extant capuchins distributed in CC belong to our  
371 widespread *Sapajus* CLADE 4 - which invaded those biomes only in the last 500 kyr,  
372 and have recent cranial and post-cranial adaptations to drier habitats (Wright *et al.*,  
373 2015).

374 Combining information from our biogeographical and phylogeographical  
375 analyses, we find strong evidence that all of the Atlantic Forest robust capuchin species  
376 are monophyletic clades, with the exception perhaps of *S. flavius*, which was found as  
377 part of the widespread *Sapajus* ‘clade 4’ in our MRBAYES phylogeny. Major Amazonian  
378 rivers do not appear to be strong barriers for dispersal or gene flow for *Sapajus*, with up  
379 to twelve examples of dispersal across significant Amazonian rivers demonstrated  
380 through our BEAST analysis by just the sampling included in this study.

381

### 382 *Gracile capuchin biogeography and phylogenetics*

383

384 In our analysis, *Cebus* shows strong geographical divisions, suggesting a  
385 widespread ancestor diversified in Amazonia and was split by range expansion and  
386 subsequent range contraction in the Andes and Central America. The Amazonian group  
387 (*C. yuracus*, *C. unicolor*, *C. albifrons*, *C. olivaceus* and *C. kaapori*) is about the same  
388 age as the CA group (*C. capucinus*, *C. imitator*, *C. versicolor*, *C. cesarae*, *C.*  
389 *leucocephalus* and *C. brunneus*). The recently discovered *Cebus kaapori* was initially  
390 considered to be a subspecies of *C. olivaceus* (Harada & Ferrari, 1996), but other  
391 morphological taxonomists have argued that it is a unique species (Groves 2001, 2005;  
392 Silva-Júnior, 2001), with some morphological characteristics more similar to *C.*  
393 *albifrons sensu lato* (Masterson, 1995). In our MRBAYES analysis, *Cebus kaapori* is  
394 recovered within the strongly supported *C. olivaceus* clade. A close relationship  
395 between *C. kaapori* and *C. olivaceus* makes sense, as *C. o. castaneus* is the gracile  
396 capuchin geographically most proximate to *C. kaapori*. *Cebus versicolor* is recovered as  
397 sister to *C. cesarae*. This is concordant with geographical distribution as both species  
398 are found in isolated pockets of the Magdalena Valley in Colombia. MtDNA COII



399 analysis of *Cebus albifrons sensu lato* by Ruiz-García *et al.* (2010) found *C. cesarae*  
400 and *C. pleei* as sister to *C. versicolor*.

401

#### 402 *Robust capuchin biogeography and phylogenetics*

403

404 Our phylogeographic analysis suggests robust capuchins were able to colonize  
405 an array of divergent habitat types all within a very short time period, in the last 1 Myr.  
406 Robust capuchins show evidence of multiple moves within and between all regions,  
407 except that they are absent from the Central American and Andean region. This may be  
408 explained because their arrival at the foot of the eastern Andes was so recent that the  
409 mountain range was already fully formed and impassable (Lynch Alfaro *et al.*, 2012a).

410 Consistent with this recent explosive radiation, we find little support for the  
411 internal topology of a morphologically diverse and geographically expansive clade of  
412 robust capuchins. Morphological species contained in this group include *S. libidinosus*,  
413 *S. cay*, *S. flavius*, *S. macrocephalus*, and *S. apella*. While there is significant individual  
414 variation within *Sapajus* populations, these five morphological species each display  
415 distinct morphological patterns with clear geographic correspondence (Silva-Júnior,  
416 2001), possibly as a consequence of repeated founder effects. While the recent  
417 divergence times among all the Amazonian and open region grasslands capuchins (all  
418 under 400 Ka) suggest there probably has not been time enough for speciation among  
419 these morphological types, our data do point to geographical clades that deserve further  
420 study for their morphological and behavioural population characteristics. For example,  
421 morphological differences between *S. macrocephalus* north and south of the Amazon  
422 River have already been described (Rylands *et al.*, 2013); our analysis here provides  
423 new hypotheses for geographic boundaries between major population centers within

424 Amazonian *Sapajus*, and suggests the need for new morphological and morphometric  
425 analyses for undersampled regions. We suggest nuclear genomic methods will be  
426 necessary to provide a complementary perspective to help resolve the relationships  
427 within this clade.

428         Rapid Pleistocene expansion throughout Amazonia has been recovered for the  
429 sister taxon to capuchin monkeys, the squirrel monkeys (Chiou *et al.*, 2011; Lynch  
430 Alfaro *et al.*, 2015a). *Sapajus* presence today in western Amazonia is explained in the  
431 BEAST phylogeography analysis as resulting from two distinct pathways: one was  
432 expanding from RO west across the Madeira River, into IN, and then spreading north  
433 into NE. The second was a move first from RO across the Amazon River into GU, then  
434 expansion west across the Negro River into NE, and from there south to IN. The exact  
435 distribution of *S. macrocephalus* in western Amazonia is contested (Groves, 2001,  
436 2005; Silva-Junior, 2001). Rylands *et al.* (2013) depict *S. macrocephalus* morphological  
437 diversity with Brazilian, Peruvian, Bolivian and Colombian forms; one explanation for  
438 this variation congruent with our results is that western Amazonia has been colonized  
439 multiple times from different *Sapajus* seed populations from different parts of  
440 Amazonia. In contrast, most of *Sapajus* diversity in eastern Amazonia can be explained  
441 by a single founder event from AF into RO, and another single founder event from RO  
442 into GU, with some later expansions back to RO from GU.

443         *Sapajus cay* has been the subject of controversy within robust capuchin taxonomy,  
444 and two major authorities (Groves, 2001, 2005; Silva-Junior, 2001) disagree  
445 significantly about its distribution, in part due to differences in the localities of samples  
446 available in each of their studies. *Sapajus cay* (*sensu* Rylands *et al.*, 2013) is recovered  
447 here as paraphyletic, with evidence for at least two distinct *Sapajus* populations moving  
448 independently into gallery forests in more open habitat types. *Sapajus cay* samples from

449 Paraguay, Rondônia, and Mato Grosso cluster with *S. apella* from Rondônia; while *S.*  
450 *cay* from northern central Mato Grosso forms a clade with nearby *S. apella* from Alta  
451 Floresta. These two populations have apparently converged morphologically to a lighter  
452 pelage phenotype in the drier, open regions. *Sapajus libidinosus* (*sensu* Rylands *et al.*,  
453 2013) marks a third expansion into CC from Amazonian *Sapajus*, with a resultant  
454 lighter pelage (Figure 2). All expansions into drier habitats appear to have occurred in  
455 the last 200-100 kyr, similar to findings for other Neotropical primates (Lynch Alfaro *et*  
456 *al.*, 2015b).

457         We expected to recover *Sapajus flavius* as sister to *S. xanthosternos* because of  
458 geographical proximity or sister to *S. libidinosus* based on a gradation of morphology  
459 between the two morphotypes (Silva, 2010); or possibly sister to all other *Sapajus*  
460 species, based on its unusual morphology (Lynch Alfaro *et al.*, 2012a). In our BEAST  
461 phylogeography and MRBAYES analyses, *Sapajus flavius* is recovered as part of the  
462 widespread *Sapajus* CLADE 4 (Figure 2 and 3b). However, in the symmetric BEAST  
463 phylogeography analysis, *S. flavius* is recovered as the sister group to *Sapajus* CLADE  
464 4 (Figure S2 in Appendix S3), suggesting all four Atlantic Forest *Sapajus* taxa may  
465 have diverged from each other first prior to *Sapajus* expansion into other regions. Given  
466 the difference in topology and the range of support values among analyses, the  
467 phylogenetic placement of *S. flavius* remains uncertain, though close phylogenetic  
468 affinity to *Sapajus* CLADE 4 is clear.

469  
470 *Sympatry between robust and gracile capuchins*

471

472         Overall, our data and analyses support the "Reinvasion of Amazonia" hypothesis  
473 (Lynch Alfaro *et al.*, 2012a) by *Sapajus* as indicated by the ancestral reconstructions of

474 the crown *Cebus* and crown *Sapajus* clades. The two capuchin genera diversify in  
475 isolation, with *Sapajus* species only later invading a *Cebus*-occupied Amazonia at about  
476 500 kya. The BEAST phylogeography analysis shows *Sapajus* invasion of Amazonia  
477 occurring first in Rondonia (south-western Amazon) and quickly spreading to all four  
478 quadrants of Amazonia. The recovered timing of entry into sympatry for gracile and  
479 robust capuchins indicate an event in Amazonia less than 1 Ma, as a result of *Sapajus*  
480 counter-invasion into a region already occupied by *Cebus*.

481

#### 482 ACKNOWLEDGEMENTS

483

484 Thanks to Robert Whittaker for inviting us to contribute this paper, and to Alexandre  
485 Antonelli, Marcelo Tejedor and anonymous reviewers for their excellent suggestions  
486 that have improved this article substantially. Special thanks to L.E. Olson and K. Fales,  
487 University of Alaska Museum. Support to M.G.M.L.'s PhD research was provided by a  
488 CNPq PhD fellowship (142141/2012-7) and CNPq SWE fellowship (201172/2014-3).  
489 Funding for this research was provided in part by NSF BCS 0833375 to J.W.L.A. Some  
490 of the molecular analyses and field expeditions were funded by CNPq/FAPEAM  
491 SISBIOTA Program (563348/2010-0) and CAPES (AUX n° 3261/2013) to I.P.F. and  
492 CNPq (grants ‘‘INCT em Biodiversidade e Uso da Terra da Amazônia’’ 574008/2008-  
493 0; and 310880/2012-2) and NSF-FAPESP (grant 1241066 - Dimensions US-BIOTA-  
494 São Paulo: Assembly and evolution of the Amazonian biota and its environment: an  
495 integrated approach) to A.A. This is article number ### of the Amazonian Mammals  
496 Research Group.

497

#### 498 REFERENCES

499

500 Aleixo, A. & Rossetti, D.D.F. (2007) Avian gene trees, landscape evolution, and  
501 geology: towards a modern synthesis of Amazonian historical biogeography?

502 *Journal of Ornithology*, **148**, 443-453.

503 Baele, G., Lemey, P., Bedford, T., Rambaut, A., Suchard, M.A. & Alekseyenko, A.V.

504 (2012) Improving the accuracy of demographic and molecular clock model

505 comparison while accommodating phylogenetic uncertainty. *Molecular Biology*

506 *and Evolution*, **29**, 2157-2167.

507 Baele, G., Li, W.L.S., Drummond, A.J., Suchard, M.A. & Lemey, P. (2013) Accurate

508 model selection of relaxed molecular clocks in Bayesian phylogenetics. *Molecular*

509 *Biology and Evolution*, **30**, 239-243.

510 Boubli, J.P., Rylands, A.B., Farias, I.P., Alfaro, M.E. & Lynch Alfaro, J. (2012) *Cebus*

511 phylogenetic relationships: a preliminary reassessment of the diversity of the

512 untufted capuchin monkeys. *American Journal of Primatology*, **74**, 381-393.

513 Boubli, J.P., Ribas, C., Lynch Alfaro, J.W., Alfaro, M.E., Silva, M.N.F., Pinho, G.M.,

514 & Farias, I.P. (2015) Spatial and temporal patterns of diversification on the

515 Amazon: A test of the riverine hypothesis for all diurnal primates of Rio Negro

516 and Rio Branco in Brazil. *Molecular Phylogenetics and Evolution*, **82**, 400-412.

517 Buckner, J.C., Lynch Alfaro, J., Rylands, A.B. & Alfaro, M.E. (2015) Biogeography of

518 the marmosets and tamarins (Callitrichidae). *Molecular Phylogenetics and*

519 *Evolution*, **82**, 413-425.

520 Chiou, K.L., Pozzi, L., Lynch Alfaro, J.W. & Di Fiore, A. (2011) Pleistocene

521 diversification of living squirrel monkeys (*Saimiri* spp.) inferred from complete

522 mitochondrial genome sequences. *Molecular Phylogenetics and Evolution*, **59**,

523 736-745.

524 Cracraft, J. (1985) Historical biogeography and patterns of differentiation within the  
525 South American avifauna: areas of endemism. *Ornithological Monographs*, **36**,  
526 49-84.

527 Drummond, A.J., Suchard, M.A., Xie, D. & Rambaut, A. (2012) Bayesian  
528 phylogenetics with BEAUti and the BEAST 1.7 *Molecular Biology and Evolution*,  
529 **29**, 1969-1973.

530 Groves, C.P. (2001) *Primate taxonomy*. Smithsonian Institution Press, Washington, DC.

531 Groves, C.P. (2005) Order Primates. *Mammal Species of the World: A Taxonomic and*  
532 *Geographic Reference* (ed. by Wilson, D.E. & Reeder, D.M.), pp 111–184, Johns  
533 Hopkins University Press, Baltimore, MD.

534 Harada, M.L. & Ferrari, S.F. (1996) Reclassification of *Cebus kaapori* Queiroz 1992  
535 based on new specimens from eastern Pará, Brazil. XVIth Congress of the  
536 International Primatological Society, XIXth Congress of the American Society of  
537 Primatologists. Madison, Wisconsin, USA.

538 Jameson Kiesling, N.M., Yi, S.V., Xu, K., Sperone, F.G. & Wildman, D. (2015) The  
539 tempo and mode of New World monkey evolution and biogeography in the  
540 context of phylogenomic analysis. *Molecular Phylogenetics and Evolution*, **82**,  
541 386-399.

542 Kay, R.F., (2015) Biogeography in deep time – what do phylogenetics, geology and  
543 paleoclimate tell us about early platyrrhine evolution? *Molecular Phylogenetics*  
544 *and Evolution*, **82**, 358-374.

545 Landis, M.J., Matzke, N.J., Moore, B.R. & Huelsenbeck, J.P. (2013) Bayesian analysis  
546 of biogeography when the number of areas is large. *Systematic Biology*, **62**, 789-  
547 804.

548 Lynch Alfaro, J.W., Boubli, J.P., Olson, L.E., Di Fiore, A., Wilson, B., Gutierrez-  
549 Espeleta, G.A., Chiou, K.L., Schulte, M., Neitzel, S., Ross, V., Schwochow, D.,  
550 Farias, I., Janson, C. & Alfaro, M.E. (2012a) Explosive Pleistocene range  
551 expansion leads to widespread Amazonian sympatry between robust and gracile  
552 capuchin monkeys. *Journal of Biogeography*, **39**, 272-288.

553 Lynch Alfaro, J.W., Silva-Júnior, J.S. & Rylands, A.R. (2012b) How different are  
554 robust and gracile capuchin monkeys? An argument for the use of *Sapajus* and  
555 *Cebus*. *American Journal of Primatology*, **74**, 273-286.

556 Lynch Alfaro, J.W., Izar, P. & Ferreira, R.G. (2014) Capuchin monkey research  
557 priorities and urgent issues. *American Journal of Primatology*, **76**, 705-720.

558 Lynch Alfaro, J.W., Boubli, J.P., Paim, F.P., Ribas, C.C., da Silva, M.N.F., Messias,  
559 M., Röhe, F., Mercês, M.P., Silva Júnior, J.S., Silva, C.R., Pinho, G.M.,  
560 Koshkarian, G., Nguyen, M.T.T., Harada, M.L., Rabelo, R.M., Queiroz, H.L.,  
561 Alfaro, M.E. & Farias, I.P. (2015a) Biogeography of squirrel monkeys (genus  
562 *Saimiri*): South-central Amazon origin and rapid pan-Amazonian diversification  
563 of a lowland primate. *Molecular Phylogenetics and Evolution*, **82**, 436-454.

564 Lynch Alfaro, J.W., Cortés-Ortiz, L., Di Fiore, A. & Boubli, J.P. (2015b) Special issue:  
565 Comparative biogeography of Neotropical primates. *Molecular Phylogenetics and*  
566 *Evolution*, **82**, 518-529.

567 Masterson T.J. (1995) Morphological relationships between the Ka'apor capuchin  
568 (*Cebus kaapori* Queiroz 1992) and other male *Cebus* crania: a preliminary report.  
569 *Neotropical Primates*, **3**, 165-171.

570 Martins Jr., A.M.G., Amorim, N., Carneiro, J.C., Affonso, P.R.A.M., Sampaio, I. &  
571 Schneider, H. (2014) *Alu* elements and the phylogeny of capuchin (*Cebus* and  
572 *Sapajus*) monkeys. *American Journal of Primatology*, **77**, 368-375.

573 Matthews, L.J. (2012) Variations in sexual behavior among capuchin monkeys function  
574 for conspecific mate recognition; a phylogenetic analysis and a new hypothesis for  
575 female proceptivity in tufted capuchins. *American Journal of Primatology*, **74**,  
576 287-298.

577 Matzke, N. (2013) 'BioGeoBEARS': BioGeography with Bayesian (and Likelihood)  
578 Evolutionary Analysis in R Scripts. University of California, Berkeley, Berkeley,  
579 CA. R package, version 0.2.1, published July 27, 2013. <[http://CRAN.R-](http://CRAN.R-project.org/package=BioGeoBEARS)  
580 [project.org/package=BioGeoBEARS](http://CRAN.R-project.org/package=BioGeoBEARS)>.

581 Matzke, N.J. (2014) Model selection in historical biogeography reveals that founder-  
582 event speciation is a crucial process in Island Clades. *Systematic Biology*, **63**, 951-  
583 970.

584 Mercês, M.P., Lynch Alfaro, J.W., Ferreira, W.A.S., Harada, M.L. & Silva Júnior, J.S.  
585 (2015) Morphology and mitochondrial phylogenetics reveal that the Amazon  
586 River separates two eastern squirrel monkey species: *Saimiri sciureus* and *S.*  
587 *collinsi*. *Molecular Phylogenetics and Evolution*, **82**, 426-435.

588 Nascimento, F., Lazar, A., Seuánez, H & Bonvicino, C. (2015) Reanalysis of the  
589 biogeographical hypothesis of range expansion between robust and gracile  
590 capuchin monkeys. *Journal of Biogeography*, **42**, 1349-1363.

591 Oliveira, S.G., Lynch Alfaro, J.W. & Veiga, L.M. (2014) Activity budget, diet and  
592 habitat use in the critically endangered Ka'apor capuchin monkey (*Cebus kaapori*)  
593 in Pará State, Brazil: A preliminary comparison to other capuchin monkeys.  
594 *American Journal of Primatology*, **76**, 919-931.

595 Queiroz, H.L. (1992) A new species of capuchin monkey, genus *Cebus* Erxleben 1977  
596 (Cebidae, Primates), from eastern Brazilian Amazonia. *Goeldiana Zoologia*, **15**, 1-  
597 3.



598 Perelman, P., Johnson, W.E., Roos, C., Seuánez, H.N., Horvath, J.E., Moreira, M.A.  
599 M., Kessing, B., Pontius, J., Roelke, M., Rumpler, Y., Schneider, M.P.C., Silva,  
600 A., O'Brien, S.J., & Pecon-Slattery, J. (2011) A molecular phylogeny of living  
601 primates. *PLoS Genetics*, **7**, e1001342.

602 Ree, R.H., Moore, B.R., Webb, C.O. & Donoghue, M.J. (2005) A likelihood framework  
603 for inferring the evolution of geographic range on phylogenetic trees. *Evolution*,  
604 **59**, 2299-2311.

605 Ree, R.H. & Smith, S.A. (2008) Maximum-likelihood Inference of Geographic Range  
606 Evolution by Dispersal, Local Extinction, and Cladogenesis. *Systematic Biology*,  
607 **57**, 4-14.

608 Robins, J.H., Tintinger, V., Aplin, K.P., Hingston, M., Matisoo-Smith, E., Penny, D. &  
609 Lavery, S.D. (2014) Phylogenetic species identification in *Rattus* highlights rapid  
610 radiation and morphological similarity of New Guinean species. *PlosOne*, **9**,  
611 e98002.

612 Ronquist, F. (1997) Dispersal-vicariance analysis: A new approach to the quantification  
613 of historical biogeography. *Systematic Biology*, **46**, 195-203.

614 Ronquist, F., Teslenko, M., van der Mark, P., Ayres, D.L., Darling, A., Höhna, S.,  
615 Larget, B., Liu, L., Suchard, M.A. & Huelsenbeck, J.P. (2012) MRBAYES 3.2:  
616 efficient Bayesian phylogenetic inference and model choice across a large model  
617 space. *Systematic Biology*, **61**, 539–42

618 Ruiz-García, M., Castillo, M.I., Vásquez, C., Rodríguez, K., Pinedo, M., Shostell, J. &  
619 Leguizamon, N. (2010) Molecular phylogenetics and phylogeography of the  
620 white-fronted capuchin (*Cebus albifrons*; Cebidae, Primates) by means of mtCOII  
621 gene sequences. *Molecular Phylogenetics and Evolution*, **57**, 1049-1061.

- 622 Ruiz-García, M., Castillo, M.I., Lichilín-Ortiz, N. & Pinedo-Castro, M. (2012)  
623 Molecular relationships and classification of several tufted capuchin lineages  
624 (*Cebus apella*, *Cebus xanthosternos* and *Cebus nigritus*, Cebidae), by means of  
625 mitochondrial Cytochrome Oxidase II gene sequence. *Folia Primatologica*, **83**,  
626 100-125.
- 627 Ruiz-García, M., Castillo, M.I. & Luengas-Villamil, K. (2016) Is it misleading to use  
628 *Sapajus* (robust capuchins) as a genus? A review of the evolution of the capuchins  
629 and suggestions on their systematics. *Phylogeny, Molecular Population Genetics,*  
630 *Evolutionary Biology and Conservation of the Neotropical Primates*. Ed. M. Ruiz-  
631 Garcia & J. M. Shostell, Nova Science Publisher Inc., New York.
- 632 Rylands, A.B., Mittermeier, R.A., Bezerra, B.M., Paim, F.P. & Queiroz, H.L. (2013)  
633 Species accounts of Cebidae. *Handbook of the Mammals of the World: vol. 3.*  
634 *Primates* (ed. by Mittermeier, R.A., Rylands, A.B. and Wilson, D.E.), pp. 390-  
635 413, Barcelona: Lynx Edicions.
- 636
- 637 Schneider, H. & Sampaio, I. (2015) The systematics and evolution of New World  
638 primates--A review. *Molecular Phylogenetics and Evolution*, **82**, 348-357.
- 639 Silva, T.C.F. (2010) Estudo da variação na pelagem e da distribuição geográfica em  
640 *Cebus flavius* e *Cebus libidinosus* do Nordeste do Brasil. MSc Dissertation.  
641 Programa de Pós-Graduação em Ciências Biológicas – Zoologia. 41p.
- 642 Silva-Júnior, J.S. (2001) *Especiação nos macacos-prego e caiararas, gênero Cebus*  
643 *Erleben, 1777 (Primates, Cebidae)*. PhD Thesis, Universidade Federal do Rio de  
644 Janeiro, Rio de Janeiro, Brazil.
- 645 Simon, M.F., Grether, R., Queiroz, L.P., Skema, C., Pennington, R.T. & Hughes, C.E.  
646 (2009) Recent assembly of the Cerrado, a Neotropical plant diversity hotspot, by

647 in situ evolution of adaptations to fire. *Proceedings of the National Academy of*  
648 *Sciences USA*, **106**, 20359-20364.

649 Springer, M.S., Meredith, R.W., Gatesy, J., Emerling, C.A., Park, J., Rabosky, D.L.,  
650 Stadler, T., Steiner, C., Ryder, O.A., Janecka, J.E., Fisher, C.A. & Murphy, W.J.  
651 (2012) Macroevolutionary dynamics and historical biogeography of primate  
652 diversification inferred from a species supermatrix. *PlosOne*, **7**, e49521.

653 Tobe, S.S., Kitchener, A.C. & Linacre, A.M.T. (2010) Reconstructing mammalian  
654 phylogenies: a detailed comparison of the Cytochrome *b* and Cytochrome Oxidase  
655 Subunit 1 mitochondrial genes. *PlosOne*, **5**, e14156.

656 Wallace, A.R. (1852) On the monkeys of the Amazon. *Proceedings of the Zoological*  
657 *Society of London*, **20**, 107-110.

658 Ward, R.D., Zemlak, T.S., Innes, B.H., Last, P.R. & Hebert, P.D.N. (2005) DNA  
659 barcoding Australia's fish species. *Philosophical Transactions of the Royal Society*  
660 *B: Biological Sciences*, **360**, 1847-1857.

661 Wright, K.A., Wright, B.W., Ford, S.M., Fragaszy, D., Izar, P., Norconk, M.,  
662 Masterson, T., Hobbs, D.G., Alfaro, M.E. & Lynch Alfaro, J.W. (2015) The  
663 effects of ecology and evolutionary history on robust capuchin morphological  
664 diversity. *Molecular Phylogenetics and Evolution*, **82**, 455-466.

665 Zhang, Y., Zhang, H., Gao, T. & Miao, Z. (2011) Structure of mitochondrial DNA  
666 control region and molecular phylogenetic relationship among three founders of  
667 genus *Pleuronectes*. *Biochemical Systematics and Ecology*, **39**, 627-634.

668

#### 669 SUPPORTING INFORMATION

670 Additional Supporting Information may be found in the online version of this article:

671 **Appendix S1** Details of individual and GenBank accession numbers.

672 **Appendix S2** Supplementary details of methods.

673 **Appendix S3** Supporting results.

674

675 **BIOSKETCH:**

676

677 **Marcela G. M. Lima** is a biologist interested in biogeography, phylogenetics and  
678 population genetics, with a focus on Neotropical vertebrates. Author Contributions:

679 M.G.M.L., J.C.B., M.E.A. and J.W.L.A. designed and ran phylogenetic and

680 biogeographic analyses and developed the article; M.G.M.L., J.S.S.J., A.M., A.L.,

681 I.P.F., J.P.B., F.R., H.Q., M.N.S., A.D.F. and J.W.L.A. collected and curated sample

682 material; M.G.M.L., A.M. and I.P.F. generated new genetic sequences; K.C.,

683 M.G.M.L., J.C.B. and J.L.A. designed maps and figures; and all authors contributed to

684 the writing of the article.

685

686 Editor: Alexandre Antonelli

687

688 **Tables**

689

690 **Table 1** Comparison between models estimated in ‘BioGeoBEARS’. For each model

691 implemented in the analysis are shown: values of log-likelihood (LnL), numbers of

692 parameters and Akaike’s information criteria (AIC,  $\Delta$ AIC and AICc Weight).

693

<b>Model</b>	<b>LnL</b>	<b>No. Parameters</b>	<b>AICc</b>	<b><math>\Delta</math>AICc</b>	<b>AICc Weight</b>
DEC	-17.29	2	39.92	0	0.68
DIVALIKE	-18.9	2	43.14	3.22	0.14
BAYAREALIKE	-21.41	3	51.81	11.9	0
DEC+J	-17.19	3	43.38	3.46	0.12
DIVALIKE+J	-18.61	3	46.22	6.3	0.03
BAYAREALIKE+J	-18.61	3	46.22	6.3	0.03

694 **Figure Legends**

695

696 **Figure 1** Maps of capuchin monkey sample provenance: (a) *Cebus*, (b) *Sapajus*.

697 **Figure 2** Asymmetric discrete states time tree for the widespread clade of robust  
698 capuchin monkeys with ancestral state reconstruction for regions of ancestral nodes and

699 map delineating centres of endemism used for the BEAST phylogeography analyses:

700 GU: Guianas; NE: Negro; RO: Rondonia; IN: Inambari; CC: Caatinga, Cerrado and

701 Central Grasslands; AF: Atlantic Forest. Branches are colour-coded to regions from

702 inset map. Posterior probabilities are shown as asterisks for above 0.95 and as circles

703 for between 0.70 and 0.95.

704 **Figure 3** Bayesian consensus tree for capuchin phylogeny: (a) *Cebus* and *Sapajus*

705 phylogeny and (b) details of *Sapajus* CLADE 4; (c) map with minimum convex

706 polygons to show geographic distribution of major subclades within the widespread

707 *Sapajus* clade. Subclades are presented as minimum convex polygons that connect

708 sample localities. Branches are colour-coded to regions from polygons map. Posterior

709 probabilities are shown as asterisks for above 0.95 and as circles for between 0.70 and

710 0.95. Sample species ID and number correspond to Table S1 in Appendix S1.

711 **Figure 4** 'BioGeoBEARS' ancestral range reconstruction of capuchin monkeys using

712 the DEC model. CA = Central America/Andes; AM = Amazonia; CC = Caatinga,

713 Cerrado and Central Grasslands; AF = Atlantic Forest. Inset map for four regions of

714 analysis.

715