



## Novel RNA Virus Genome Discovered in Ghost Ants (*Tapinoma melanocephalum*) from Hawaii

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**ABSTRACT** Here, we report the full-genome sequence of Milolii virus, a novel singlestranded (positive-sense) RNA virus discovered from *Tapinoma melanocephalum* ants in Hawaii. The genome is 10,475 nucleotides long, encoding a polyprotein of 3,304 amino acids.

The ghost ant (*Tapinoma melanocephalum* [Fabricius]) is a widely distributed invasive pest, probably of African or Asian origin (1), that is now found in tropical and subtropical climates as well as in glass houses in temperate regions (2). The species was first recorded in Hawaii in 1899 (3) and soon became a common household pest (4).

Ghost ants are now a major pest across the Pacific Islands (5) and Florida (6) and are listed in the Global Invasive Species Database (7). They are highly adaptable, feeding on waste food (2), and in hospitals, they pose an additional threat of carrying pathogenic bacteria (8).

The combination of an invasive species with an emerging disease poses a potentially major risk to biodiversity (9), since many RNA viral pathogens have wide host ranges that can belong to different but overlapping food webs. This provides vast potential for viral pathogens to spread via different networks to new hosts (10, 11).

RNA was extracted from two pooled samples of 20 to 25 *T. melanocephalum* ants collected from inside two honey bee (*Apis mellifera*) hives from an apiary in Milolii, Big Island, Hawaii, in December 2012, using the RNeasy minikit (Qiagen). Oligo(dT) priming was used to create cDNA libraries, which were sequenced using the Illumina HiSeq 2000 platform at the Earlham Institute, Norwich, United Kingdom (formerly the Genome Analysis Centre), to produce 100-bp paired-end reads. Quality filtering was carried out using FastQC. Kontamination, a pipeline developed by TGAC to remove host reads, was applied using the related reference *Linepithema humile* ant genome due to the unavailability of a *T. melanocephalum* genome. The resulting reads were pooled and IVA version 1.0.3 (12) was used to generate *de novo* assembled contigs, which resulted in the assembly of the Milolii virus genome.

The genome is 10,475 nucleotides (nt) long with a 3' polyadenylated tail end and a 9,930-nt open reading frame encoding a polyprotein of 3,304 amino acids. A BLASTx search of the protein-coding sequences against the NCBI protein database revealed conserved RNA helicase and RNA-dependent RNA polymerase (RdRp) domains typical of single-stranded (positive) RNA viruses. The protein-coding sequences showed closest matches (29% to 30%) to the hypothetical protein of Hubei Tetragnatha maxillosa virus 5 (13) and the P1 protein of Acrythosiphon pisum virus (14), both invertebrate viruses. The coverages across the protein-coding sequences were 56% and 60%, respectively. For each sample, total reads (FASTA format) were mapped against the Milolii virus genome in Geneious (Biomatters). Both showed full-length genome coverages with depths ranging from approximately  $1,000 \times$  to  $1,000,000 \times$  (data not shown), with 28.12% and 69.65% of the total reads aligning to Milolii virus. Furthermore, full-length

Received 25 May 2017 Accepted 12 June 2017 Published 27 July 2017

Citation Brettell LE, Mordecai GJ, Pachori P, Martin SJ. 2017. Novel RNA virus genome discovered in ghost ants (*Tapinoma melanocephalum*) from Hawaii. Genome Announc 5:e00669-17. https://doi.org/10.1128/ genomeA.00669-17.

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Address correspondence to Laura E. Brettell, l.e.brettell@edu.salford.ac.uk.

\* Present address: Gideon J. Mordecai, Department of Earth, Ocean and Atmospheric Sciences, University of British Colombia, Vancouver, Canada. genomes were assembled individually from each of the two samples, and Geneious nucleotide alignments revealed that the sequences share 99.9% pairwise nucleotide similarity.

**Accession number(s).** This full-genome sequence has been submitted to GenBank under the accession number MF155030.

## ACKNOWLEDGMENTS

We thank the C. B. Dennis British Beekeepers' Research Trust for funding this research. L.E.B. and S.J.M. are funded by the C. B. Dennis and Apis M; L.E.B. also receives funding from the British Beekeepers' Association. G.J.M. was funded by the British Beekeepers' Association and the University of Reading at the time this study was conducted.

Additional thanks go to Ethel Villalobos and Scott Nikaido for their assistance with arranging sampling in Hawaii.

## REFERENCES

- 1. Wheeler WM. 1910. Ants, their structure, development and behavior. Columbia University Press, New York, NY.
- Smith MR. 1965. House-infesting ants of the Eastern United States: their recognition, biology, and economic importance. USDA Technical Bulletin 1326:1–105. U.S. Department of Agriculture, Washington, DC.
- 3. Forel A. 1899. Fauna Hawaiiensis, vol 1: Heterogyna (Formicidae). Cambridge University Press, Cambridge, UK.
- 4. Clagg CF. 1957. Notes and exhibitions: *Tapinoma melanocephalum*. Proc Hawaii Entomol Soc 16:197.
- 5. O'Connor S. 2004. Pacific Ant Protection Plan (PAPP). Aliens 19/20:14.
- Klotz JH, Mangold JR, Vail KM, Davis LR, Jr, Patterson RS. 1995. A survey of the urban pest ants (*Hymenoptera: Formicidae*) of Peninsular Florida. Florida Entomologist 78:109–118. https://doi.org/10.2307/3495674.
- Global Invasive Species Database (GISD). 2015. Species profile: Tapinoma melanocephalum. http://www.iucngisd.org/gisd/species.php?sc=959.
- Moreira DDO, Morais V, Vieira-da-Motta O, Campos-Farinha AEDC, Tonhasca A, Jr. 2005. Ants as carriers of antibiotic-resistant bacteria in hospitals. Neotrop Entomol 34:999–1006. https://doi.org/10.1590/S1519 -566X2005000600017.
- 9. Strauss A, White A, Boots M. 2012. Invading with biological weapons: the

importance of disease-mediated invasions. Funct Ecol 26:1249–1261. https://doi.org/10.1111/1365-2435.12011.

- McMahon DP, Fürst MA, Caspar J, Theodorou P, Brown MJ, Paxton RJ. 2015. A sting in the spit: widespread cross-infection of multiple RNA viruses across wild and managed bees. J Anim Ecol 84:615–624. https:// doi.org/10.1111/1365-2656.12345.
- Mordecai GJ, Brettell LE, Pachori P, Villalobos EM, Martin SJ, Jones IM, Schroeder DC. 2016. Moku virus; a new *lflavirus* found in wasps, honey bees and varroa. Sci Rep 6:34983. https://doi.org/10.1038/srep34983.
- Hunt M, Gall A, Ong SH, Brener J, Ferns B, Goulder P, Nastouli E, Keane JA, Kellam P, Otto TD. 2015. IVA: accurate *de novo* assembly of RNA virus genomes. Bioinformatics 31:2374–2376. https://doi.org/10.1093/ bioinformatics/btv120.
- Shi M, Lin XD, Vasilakis N, Tian JH, Li CX, Chen LJ, Eastwood G, Diao XN, Chen MH, Chen X, Qin XC, Widen SG, Wood TG, Tesh RB, Xu J, Holmes EC, Zhang YZ. 2015. Divergent viruses discovered in arthropods and vertebrates revise the evolutionary history of the *Flaviviridae* and related viruses. J Virol 90:659–669. https://doi.org/10.1128/JVI.02036-15.
- Van der Wilk F, Dullemans AM, Verbeek M, Van den Heuvel JFJM. 1997. Nucleotide sequence and genomic organization of Acyrthosiphon pisum virus. Virology 238:353–362. https://doi.org/10.1006/viro.1997.8835.