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Changes in tendon spatial frequency parameters with loading

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22 **Abstract**

23 To examine and compare the loading related changes in micro-morphology of the patellar tendon.
24 Fifteen healthy young males (age 19 ± 3 yrs, body mass 83 ± 5 kg) were utilised in a within subjects
25 matched pairs design. B mode ultrasound images were taken in the sagittal plane of the patellar tendon
26 at rest with the knee at 90° flexion. Repeat images were taken whilst the subjects were carrying out
27 maximal voluntary isometric contractions.

28 Spatial frequency parameters related to the tendon morphology were determined within regions of
29 interest (ROI) from the B mode images at rest and during isometric contractions.

30 A number of spatial parameters were observed to be significantly different between resting and
31 contracted images (Peak spatial frequency radius (PSFR), axis ratio, spatial Q-factor, PSFR amplitude
32 ratio, and the sum). These spatial frequency parameters were indicative of acute alterations in the
33 tendon micro-morphology with loading.

34 Acute loading modifies the micro-morphology of the tendon, as observed via spatial frequency analysis.
35 Further research is warranted to explore its utility with regard to different loading induced micro-
36 morphological alterations, as these could give valuable insight not only to aid strengthening of this
37 tissue but also optimization of recovery from injury and treatment of conditions such as tendinopathies.

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40 **Introduction**

41 Tendons are made up predominantly of collagen (60-85% of dry weight) (Józsa et al., 1989), with type I
42 collagen being the predominant type. There are also elastin elements 1 - 2 % (Kirkendall and Garrett,
43 1997), which are embedded with the collagen in a proteoglycan - water matrix. These elements give
44 tendons viscoelastic properties and as such they respond acutely to loading in a load (elastic) and time
45 (viscous) dependent way (Ker et al., 2000).

46

47 The collagen component of the tendon can be seen to be the main structural element and its structure
48 reflects the loading deformation characteristics seen, with a nonlinear 'toe' (beginning of loading
49 region) region as a consequence of the 'crimp' (bunching of collagen fibres) seen in resting collagen
50 structures (Diamant et al., 1972), to a then reasonably linear elastic region reflective of the elastin
51 structures within the tendon and sliding of the tropocollagen molecules and associated stretching of the
52 triple collagen helices (Folkhard et al., 1987). There is also interstitial fluid flow seen with loading of
53 tendons, possibly representing the viscous element (Hannafin and Arnoczky, 1994). The loading of
54 tendons can result in physiologic adaptations which may be beneficial or detrimental, dependant on the
55 loading. Tendons **transfer** the load via mechanotransduction from the cellular matrix to the tendon cells,
56 resulting in biochemical responses at cellular level.

57

58 Loading protocols have previously been investigated in an attempt to identify optimal strategies for
59 adaptation and or recovery from injury and conditions such as tendinopathies. For example, a number
60 of studies have measured tendon cross-sectional area (CSA) and observed any increases after a period of
61 loading. Of those that have utilized MRI to determine changes it was reported that short term loading
62 (12 weeks and 9 weeks respectively) can result in region specific hypertrophic changes (Kongsgaard et
63 al., 2007;Seynnes et al., 2009). However other potential possibilities of adaptation are intrinsic changes

64 to the tendon structure as indicated by increases in Young's Modulus (Reeves et al., 2003;Bohm et al.,
65 2014). In the Bohm et al. study, high strain rate loading was seen to be preferential in producing tendon
66 adaptation.

67

68 The mechanisms underlying adaptation are unclear. Studies examining acute cyclic loading have shown
69 changes in transverse strain. Here reductions in the tendon thickness are evident when loaded (Wearing
70 et al., 2013). The loading associated reduction in tendon thickness is suggested to be in part due to
71 fluid transfer out of the tendon. However, acute changes in thickness during loading are also indicative
72 of alterations of the tendon architecture. Here alignment and increased density of the collagen
73 structures occurs with tensile loading (York et al., 2014). Changes in the tendon component
74 arrangements can be indirectly described by the tendon stress/strain characteristic relationship. Below
75 2% strain (the toe region) represents the "stretching-out" of crimped tendon fibrils with tensile loading.
76 This typical mechanical observation due to the 'crimped' fibril pattern can change to some degree, due
77 to the differential crimp angle and crimp length between structures. With increased loading, a linear
78 region in the stress strain curve appears (up to approx 4-5 % strain). Here the collagen fibrils begin to
79 alter their conformation and align themselves in the direction of tensile loading. These characteristic
80 alterations in the tendon micro-morphology with loading may possibly be identified with ultrasound
81 imaging (Kostyuk et al., 2004).

82

83 Visualisation of the tendon using B-mode ultrasound shows an anisotropic speckle pattern in which the
84 pattern or image texture and brightness depends on the spatial distribution of the acoustic scatters
85 within the tendon. This characteristic pattern and intensity is affected by the tendon structure and
86 alignment to the ultrasound emission waves (Kannus, 2000). For healthy tendon, the speckle pattern

87 present has a spatial frequency signature with a significant magnitude element and narrow frequency
88 bandwidth about the peak spatial frequency (Bashford et al., 2008).

89

90 Analysis of tendon pathology had been previously performed in the frequency domain (Bashford et al.,
91 2008). Previous work has shown peak spatial frequency radius (PSFR) in a tendon to be correlated with
92 tendon elasticity (Kulig et al., 2016). Tendon is made up of elastic polymers. At rest, the individual
93 polymer are more bundled, they can be stretched a finite length before breaking (Rigby et al., 1959). If
94 ultrasound frequency analysis is able to categorise tendon mechanical properties it may be useful to
95 examine whether this approach is sensitive enough to detect acute alterations in the tendon micro-
96 architecture with loading.

97

98 Thus it can be seen that the acute loading and the mode of loading may influence the tendon response
99 with acute loading. Hence this study applies the use of spatial frequency analysis of the tendon B mode
100 images to examine indicators of acute changes within the tendon structure.

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104 **Materials & Methods**

105 15 male participants all gave their written informed consent and were included in this study (age 19 ± 3
106 yrs, body mass 83 ± 5 kg). All experimentation was approved by the local ethics committee and all
107 procedures were in accordance with the world medical association Declaration of Helsinki (2013).

108 B-Mode ultrasound images (7.5 MHz 100mm linear array B-mode ultrasound probe (Mylab 70, Esaote
109 Biomedica, Italy) with a depth setting of 30mm were taken of the patellar tendon in the sagittal plane,
110 both whilst they were unloaded and during maximal voluntary isometric contractions (MVC), held for
111 approx 5 secs with the knee flexed to 90° . A familiarisation isometric contraction was performed prior to
112 the test contraction for all subjects. Figure 1 shows an example of an unloaded and loaded tendon. For
113 each image pair, a region of interest (ROI) was selected corresponding to the tendon tissue seen within
114 the image. Within the ROI, Cohens d was calculated by

115

$$116 \quad d = \frac{\bar{X}}{\sigma_X} \quad 1$$

117

118 where X is the paired differences of the samples and σ_X is the standard deviation of the paired
119 differences. P-values were estimated using a permutation test on X . A total of 10000 permutations were
120 used.

121

$$122 \quad p = \frac{\sum_{n=1}^{10000} (\bar{X}_n \geq \bar{X}_0)}{10000} \quad 2$$

123

124 Here \bar{X}_0 is the observed average difference and X_n is the n-th permutation of the observed data.

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128 Image analysis and parameter extraction

129 Spatial frequency analysis was carried out in a manner similar to Bashford and co workers 2008
130 (Bashford et al., 2008). Briefly, digitally stored images in jpeg format were imported into MATLAB where
131 all analyses were completed with custom in-house algorithms. An ROI enclosing the tendon area of
132 interest was selected by cursor (See Fig 1). Within the ROI, every possible 2mm-square kernel
133 circumscribed by the ROI was analyzed. For each kernel, eight spatial frequency parameters were
134 extracted (see Table 1). The number of kernels circumscribed by the ROI was the number of
135 observations made of each parameter.

136

137 Statistical analyses

138 All statistical analyses were carried out in SPSS (Ver 20; IBM Corp) and MATLAB (ver 8.4, Mathworks,
139 Natick, MA). SPSS was used to calculate normality of data and determined by the Shapiro Wilks test.
140 Paired Student t-tests were utilised to determine any significant differences for tendon thickness and
141 between the frequency determinants for the ROI. Alpha level was set to 0.05.

142 MATLAB was used to analyze spatial frequency parameter statistics. Paired student t-tests were utilized
143 to determine any significant differences between the spatial frequency parameters between loaded and
144 unloaded tendons. The alpha level was set to 0.05.

145

146

147 **Results**

148 Table 2 shows the paired statistics for unloaded to loaded tendons. Here only the whole tendon data is
149 shown. The parameters that met statistically significant differences are the Peak Spatial Frequency
150 Radius, Axis Ratio, Mmax%, and the Sum where tendon loading produced an increase of 0.51, 0.67, 1.21
151 and decrease of 1.04 standard deviations of the estimate respectively.

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155 **Discussion**

156 The results here show characteristic changes in the frequency analysis of the patellar tendon during
157 acute maximal isometric loading. In particular, increases in the peak spatial frequency radius, axis ratio,
158 and sum were seen with a concurrent decrease in normalized peak spatial frequency amplitude
159 (Mmax%) . To understand the change of PSFR, Axis Ratio, Mmax%, and the Sum (local intensity), it is
160 necessary to understand the natural structure of the tendon. Tendons are viscoelastic in nature due to
161 the composite elements (elastin/collagen and the proteoglycan water matrix). This viscoelastic
162 characteristic results in creep whereby elongation increases with a fixed load application, and stress
163 relaxation whereby the stress is reduced with a fixed elongation (Clatworthy et al., 1999; Ker et al.,
164 2000). If the elongation is determined during application of force it can be seen to be a curvilinear
165 relationship. Here larger elongation is seen initially, probably due to the uncrimping of the collagen
166 fibrils in the direction of the applied stress and the beginning of increased lateral packing of the fibrils
167 (Diamant et al., 1972;Kirkendall and Garrett, 1997;Misof et al., 1997). After this initial elongation there
168 appears a quite linear portion, reflective of the stretching and further alignment of the collagen helices
169 and sliding of the tropocollagen molecules (Folkhard et al., 1987). These alterations in the tendon
170 structure during loading can explain an increase in the Peak Spatial Frequency Radius which is expected
171 as the collagen fibres straighten and move slightly closer together. This same process is responsible for
172 the increase in the Axis Ratio, but here it is predominantly influenced by the fibre straightening.

173 The increase in the sum measure may be reflective of alterations in the water content of the tendon. It
174 is well known that an inflamed tendon or muscle will have higher water content and will show up darker
175 on a B-Mode ultrasound scan. This is because water will decrease the scatterer density. Thus during
176 loading it may be that water or fluid is displaced temporarily through the intracellular spaces out of the
177 tendon (Hannafin and Arnoczky, 1994;Lanir et al., 1988), along with increased density/alignment of the
178 collagen fibrils, resulting in a brighter image signature and hence increased sum value. Although it must

179 be stated that the likelihood of the suggested exchange of fluid between the interstitial spaces is
180 currently unknown during short term contractions such as here and is in need of further investigation.

181

182 In addition to the loaded tendon possibly increasing the overall density of the scatterers and hence the
183 observed increased brightness, the brighter speckle pattern is proposed based on the understanding of
184 its statistics. Initially, in a relaxed state the tendon extracellular matrix (ECM) is bundled with random
185 orientations, upon elongation the random kinks in the ECM become smoother and their scattering will
186 be coherent with the nearby ECM scattering. The probability density function for the speckle pattern of
187 the tendon, which is already a Rician distribution, becomes a stronger Rician distribution moving further
188 away from the standard Rayleigh distribution which defines the speckle pattern of unstructured tissue.
189 This explanation for the increase in speckle pattern brightness agrees with the explanation showing an
190 increase of the Peak Spatial Frequency and the Axis Ratio.

191

192 Tendons that are routinely 'loaded' become stiffer and show intrinsic modifications indicative of the
193 strengthening of the tendon as it adapts to the loads experienced (Wiesinger et al., 2015). Previous
194 work has indicated these increases in stiffness with exercise (Onambélé et al., 2008; Burgess et al., 2009).
195 In contrast, unloading can be seen to result in decrease in the tendon mechanical properties (de Boer et
196 al., 2007). Recently it has been shown that the tendon mechanical properties are related to the spectral
197 frequencies in the ultrasound image of degenerative tendons (Kulig et al., 2016). Here it was stated that
198 the peak spatial frequency radius showed a good relationship with the determined stiffness of the
199 Achilles tendon ($r = 0.74$). This relationship was not seen in healthy tendons however. It was not clear
200 from the study if the spatial parameters were determined with the tendon at rest or during loading. This
201 may be of importance as healthy tendon may not react the same to loading as degenerative tendon and
202 if spectral parameters had been determined both at rest and during loading, perhaps more insight could

203 have been gained into the differences. Indeed here we show in healthy tendons using the same
204 approach, qualitative changes when going from unloaded to loaded as identified by the spectral
205 parameters. This approach could yield interesting information regarding the tendon adaptability to both
206 acute and chronic loading protocols. The ability to monitor tendon in this way may enable an index of
207 healthy tendon to be established, which may be able to detect when a tendon is beginning to maladapt.
208 Whilst we acknowledge that more work needs to be done to further validate this method i.e.
209 repeatability of measures and examination of the sensitivity under a variety of conditions to determine
210 its viability and limitations.

211 The potential information from this technique would allow changes to the training programme to be
212 made for instance in the case of an athlete. Where rehabilitation was being carried out it may give
213 valuable insight into the significance or effectiveness of the rehabilitation programme enabling
214 optimisation of the rehabilitation on an individual level.

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290 **Figure caption**

291

292 Figure 1. Typical saggital plane ultrasound image of the patellar tendon showing the enclosed ROI

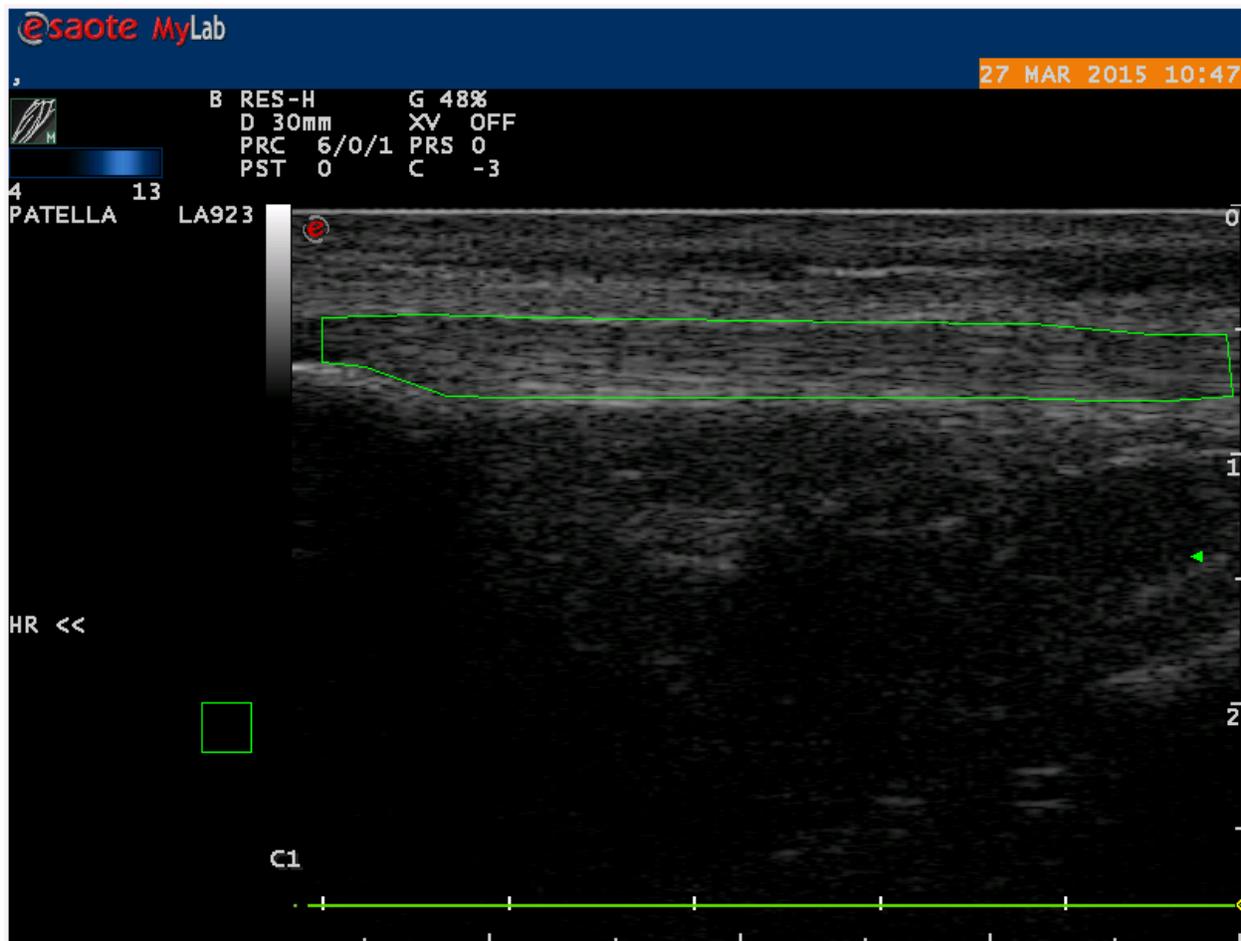
293 rectangle box in green overlaying the patellar tendon.

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296 Figures

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304 Tables

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306 Table 1. Spatial frequency parameters analysed for the tendon images and descriptions

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Table 1.		
Parameter #	Parameter Name	Description/tissue characteristic indicators
1	Peak Spatial Frequency	Distance from origin to spatial frequency peak of greatest amplitude on 2-D FFT spectrum The purity of fibre alignment modulated by the axial resolution of the scanner
2	P6 Width	Average of horizontal and vertical -6 dB widths of greatest amplitude spatial frequency peak on 2-D FFT spectrum Decreasing P6 indicates less obtuse angular entanglement between tendon fibres
3	Q6 Factor	Ratio of Peak Spatial Frequency to P6 Width A normalization factor to facilitate comparison of fibre packing with fibre alignment
4	Mmax	Value of the maximum point of amplitude on 2-D FFT The strength of the most prominent fibre spacing of the

		tendon
5	Mmax Percent	Ratio of Mmax to the total intensity of pixels in the 2-D FFT A comparison of fibres that are highly aligned with those aligned randomly
6	Sum	Sum of intensities of original image kernel pixels Overall brightness of the image
7	Axis Ratio*	Major-to-minor axis ratio of ellipsoidal fit 16 X 16 center pixel region of 2-D FFT Comparing the elongation of spatial frequencies in orthogonal directions – the higher this number, the more anisotropic the elongation is, indicating uniform alignment across the ROI
8	Ellipse Rotation*	Rotation from vertical axis of ellipsoid fit to 2-D FFT The angle between the transducer axis and the most prominent alignment of tendon fibres
* indicates parameter used by Tuthill ((Tuthill et al., 1999))		

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310

311 Table 2. The paired statistics for unloaded to loaded tendons. Unloaded and Loaded data are mean
 312 values. Parameters for which a significant change were observed are the Peak Spatial Frequency
 313 Radius, Axis Ratio, Mmax%, and the Sum (shown in bold).

314

Whole Tendon (Paired Stats)

	unloaded to loaded			
	Unloaded	Loaded	Cohen's d	p-value
Peak Spatial	2.195	2.282	0.51	0.03
Frequency Radius				
P6	0.640	0.639	-0.04	0.43
Q6	28.05	29.26	0.50	0.04
MMax	1458	1455	-0.08	0.38
M-Max_%	2.456	2.153	-1.04	0.0006
Axis Ratio	4.269	4.494	0.67	0.01
Ellipse Rotation	89.81	89.85	0.26	0.17
Sum	60425	67841	1.21	0.0002

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316