

Investigating Cholesterol Metabolism and Ageing Using a Systems Biology Approach

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Shortened Title: Cholesterol Metabolism and Ageing

Key Words: Cholesterol, ageing, systems biology

Abstract

Cardiovascular disease (CVD) accounted for 27% of all deaths in the United Kingdom in 2014, and was responsible for 1.7 million hospital admissions in 2013/14. This condition becomes increasingly prevalent with age, affecting 34.1 and 29.8% of males and females over 75 years of age respectively in 2011. The dysregulation of cholesterol metabolism with age, often observed as a rise in low density lipoprotein cholesterol (LDL-C), has been associated with the pathogenesis of CVD. To compound this problem, it is estimated by 2050, 22% of the world's population will be over 60 years of age, in culmination with a growing resistance and intolerance to pre-existing cholesterol regulating drugs such as statins. Therefore, it is apparent research into additional therapies for hypercholesterolaemia and CVD prevention is a growing necessity. However, it is also imperative to recognise this complex biological system cannot be studied using a reductionist approach; rather its biological uniqueness necessitates a more integrated methodology, such as that offered by systems biology. In this review we firstly discuss cholesterol metabolism and how it is affected by diet and the ageing process. Next, we describe therapeutic strategies for hypercholesterolaemia, and finally how the systems biology paradigm can be utilised to investigate how ageing interacts with complex systems such as cholesterol metabolism.

Introduction

Life expectancy has increased dramatically (**Figure 1**). In the UK, males and females born in 1982, had a life expectancy of 71.1 and 77.0 years respectively, while the projected values for 2082 are 89.7 and 92.6 years⁽¹⁾. Thus, we are witnessing a staggering demographic shift in favour of older people (**Figure 2**). For instance, it has been estimated the percentage of individuals in the UK over 60 years of age will double to 22% by 2050, when compared to 2000⁽²⁾. Remaining disease free is a significant challenge faced by older people, as the prevalence of many conditions increases with age (**Figure 3**). Of the diseases associated with advancing age, CVD is the leading cause of morbidity in individuals over 60 years of age⁽³⁾. The dysregulation of cholesterol metabolism is intimately associated with the pathogenesis of CVD⁽⁴⁾, and age-related alterations in the metabolism of cholesterol are implicated in the disturbance of this system⁽⁵⁾. These include, a decrease in LDL-C clearance; a potential increase in cholesterol absorption; a decrease in bile acid synthesis; and a decrease in bacterial bile acid modification^(6; 7; 8). It is likely these alterations play a role in the

accumulation of LDL-C, and disease pathogenesis. The accumulation of plasma cholesterol can also be moderated by diet, while pharmaceutical and pre- and probiotic administration have largely been associated with reduced LDL-C levels and CVD risk^(9; 10; 11; 12; 13).

Traditionally, when nutritionists have investigated complex metabolic pathways, such as cholesterol metabolism, they have utilised conventional wet laboratory techniques. However, studying cholesterol metabolism and its interaction with both diet and ageing using conventional approaches is challenging, due to the integrated nature of this system, and the time scales involved in studying the effects of the ageing process. Traditional *in vivo* or *in vitro* techniques can also be limited when testing a hypothesis, as such approaches can be resource-intensive, expensive, impractical and potentially unethical⁽¹⁴⁾. Thus, utilisation of the systems biology approach is becoming an increasingly important tool in nutrition based research, as systems biology overcomes a number of the challenges outlined above, but more importantly, facilitates the integration of data generated from a diverse range of sources⁽¹⁴⁾, leading to an improved understanding of how cellular dynamics influence the behaviour of tissues and ultimately the health of whole organ systems⁽¹⁵⁾. Thus, the systems biology approach seeks to understand complex biological systems by studying them in a more holistic manner, in contrast to the reductionist approach regularly adopted in human nutrition. At the core of the systems biology approach is computational modelling. Computational modelling is an abstract process that is used to represent the dynamics of a biological system in a precise manner using mathematics. The steps involved in building a computational model are outlined in **Figure 4**. Computational models are now used to model a diverse range of complex nutrient centred pathways including cholesterol metabolism for a number of reasons. Firstly, computational models are capable of providing quantitative data on the interaction of molecular components⁽¹⁶⁾. Secondly, nutrient-based interactions are inherently complex and often non-linear in nature^(17; 18; 19), and can involve complex feedback and feed-forward loops^(20; 21; 22). Thus, it is challenging and even unfeasible to reason about these by human intuition alone. Computational modelling offers an alternative means of handling this complexity, thus utilisation of computational modelling alongside experimental work provides a means of representing the dynamics of complex biological systems. Models can be used to simulate intrinsic perturbations, such as those associated with ageing and extrinsic perturbations, such as diet. Output from the model provides an overview of how these changes impact the dynamics of the system, and the implications this has for health-span. In this review we present an overview of cholesterol metabolism and discuss how ageing

impacts its regulatory mechanisms. We also discuss how diet influences cholesterol metabolism, and how the dysregulation of this system influences health. Next, we discuss therapeutic strategies for the treatment of hypercholesterolaemia, namely dietary, pharmacological, and probiotic intervention. Finally, we describe how we are using the systems biology framework to investigate cholesterol metabolism and the impact ageing has on it. Specifically, there is a focus on how we have used computational modelling and how we are exploring this approach with simulated digestive tracks.

Overview of Cholesterol Metabolism

Cholesterol plays a vital role in the body as a component of cell membranes, and precursor to steroid hormones and bile acids. Whole body cholesterol metabolism is encapsulated by cholesterol ingestion, absorption, excretion and synthesis. These factors interact in a coordinated fashion to regulate whole-body cholesterol balance, with subtle changes to individual components influencing the behaviour of the others, so that cholesterol balance is maintained. In the next sections we will outline in detail the complexities of cholesterol metabolism and how ageing interacts with it, thus emphasising the need for a systems biology approach when investigating it.

Cholesterol Ingestion and Absorption

In the UK, the average daily intake of cholesterol is 304 and 213mg for males and females respectively⁽²³⁾; 10-15% of which is in the esterified form⁽²⁴⁾. In the small intestine, esterified cholesterol is hydrolysed to form free cholesterol, which is more readily incorporated into bile acid micelles, which facilitate the absorption of cholesterol via Niemann-Pick C1-Like 1 protein (NPC1L1)^(25; 26). Additionally, phytosterols can also be absorbed via NPC1L1⁽²⁷⁾. Intestinal absorption of cholesterol and phytosterols can be limited by heterodimer ATP-binding cassette (ABC) G5/G8, which effluxes these sterols back to the intestinal lumen⁽²⁸⁾. Acyl coenzyme A: cholesterol acyltransferase 2 (ACAT-2) esterifies internalised cholesterol, which is then incorporated into a nascent chylomicron via microsomal triglyceride transfer protein (MTP)^(29; 30). The nascent chylomicron then exits the enterocyte by exocytosis into the lymphatic system before entering the blood stream⁽³¹⁾. The nascent chylomicron is converted to a mature chylomicron upon acquisition of apolipoprotein (apo) C-II and E from high density lipoproteins (HDL). Apo C-II activates lipoprotein lipase (LPL) on the capillary endothelium of adipose or muscle tissue, which in turn catalyses the hydrolysis of

triacylglycerol (TAG)^(32; 33). Apo C-II is then returned to HDL, and hepatic low density lipoprotein receptors (LDLr) and LDL receptor-related protein (LRP) recognise apo B-48 and E, initiating the absorption of the chylomicron remnants⁽³⁴⁾.

Cholesterol Synthesis

Cholesterol is synthesised endogenously in all nucleated cells in the body from acetyl CoA⁽³⁵⁾. Renfurm et al. (2004) observed endogenous cholesterol was synthesised at a rate of 9.8 ± 6.2 mg/kg/day in healthy adults with a mean age of 32 years and mean weight of 64 kg⁽³⁶⁾. This equates to 627.2 mg/day of synthesised cholesterol, a similar value to the 710 mg/day observed by Jones and Schoeller (1990). Interestingly, cholesterol consumption can influence the synthesis of endogenous cholesterol; an increase from 173 to 781 mg/day of dietary cholesterol has been observed to decrease the rate of sterol synthesis by 34%⁽³⁷⁾.

Cholesterol synthesis commences when acetoacetyl CoA thiolase catalyses the interconversion of acetyl CoA and acetoacetyl CoA. One molecule of acetyl CoA and one molecule acetoacetyl CoA undergo a condensation reaction by 3-hydroxy-3-methylglutaryl-coenzyme A (HMG CoA) synthase to form a molecule of HMG CoA. HMG CoA reductase, with the addition of 2 nicotinamide adenine dinucleotide phosphate (NADPH) molecules, then catalyse the conversion of HMG CoA to mevalonate. As the rate limiting enzyme of cholesterol biosynthesis, HMG CoA reductase is the therapeutic target of statins, for the treatment of hypercholesterolaemia, and the prevention of atherosclerosis⁽³⁸⁾. Phosphorylation of mevalonate by the enzyme mevalonate kinase forms mevalonate-5P, which undergoes further phosphorylation to form mevalonate-5PP via the enzyme phosphomevalonate kinase. Decarboxylation and dehydration by mevalonate-5PP decarboxylase creates isopentenyl-PP (IPP), and thus its isoform dimethylallyl-PP (DMAPP) via isopentenyl diphosphate delta isomerase. Farnesyl diphosphate synthase initiates the condensation of DMAPP with one molecule of IPP and NADPH to create geranyl-PP. Further condensation and the addition of another molecule of IPP and NADPH creates farnesyl-PP. Condensation of 2 farnesyl-PP molecules by squalene synthase and NADPH forms squalene, which is then converted to squalene epoxide by squalene epoxidase, NADPH, and O₂, before undergoing cyclisation by oxidosqualene cyclase to form lanosterol⁽³⁹⁾. A series of reactions, including the branching of 7-dehydrodesmosterol to either desmosterol or 7-dehydrocholesterol, both of which can then be converted to the end product cholesterol via the enzymes 24-dehydrocholesterol reductase

(DHCR24) and 7-dehydrocholesterol reductase (DHCR7) concludes the *de novo* synthesis of cholesterol^(40; 41).

Lipoprotein Dynamics and Reverse Cholesterol Transport

Very low density lipoprotein cholesterol (VLDL-C) is formed from the hepatic pool of cholesterol to transport endogenously synthesised TAG to the tissues⁽⁴²⁾. Partial hydrolysis of VLDL by LPL forms intermediate density lipoprotein (IDL), with subsequent hydrolysis of IDL by hepatic lipase (HL) forming LDL, which acts to deliver cholesterol to the peripheral tissue⁽⁴²⁾. VLDL-C, IDL-C and LDL-C can be removed from the circulation by hepatic LDLr, while LDL-C can also be absorbed independently^(43; 44). Reverse cholesterol transport (RCT) transfers cholesterol from the tissues to the liver via HDL, reducing the risk of cholesterol accumulation and atherosclerosis⁽⁴⁵⁾. Cholesterol can be effluxed from the tissues by the receptors ABC-A1, and scavenger receptor class B member 1 (SR-B1), or via receptor independent passive diffusion to nascent HDL^(46; 47; 48). The incorporated cholesterol is then esterified by lecithin-cholesterol acyltransferase (LCAT)⁽⁴⁹⁾. Cholesterol enters the liver either directly, via the receptor SR-B1, or via the enzyme cholesteryl ester transfer protein (CETP). CETP mediates the 1:1 exchange of cholesterol from HDL with TAG from VLDL and LDL⁽⁵⁰⁾. Once in the liver, cholesterol can be removed from the body.

Cholesterol Excretion

Cholesterol can be removed from the body by two mechanisms, directly via the hepatic ABCG5/G8 receptor and effluxed to the gall bladder or alternatively, cholesterol can be converted to bile acids for faecal excretion^(51; 52). Approximately 98% of bile acids are conjugated to either taurine or glycine, as conjugation increases polarity, which reduces passive transport from the intestinal lumen into enterocytes, and allows the movement of bile acids to be tightly regulated, and under receptor control; in addition to improving solubility⁽⁵³⁾. Removal of the amino acid from conjugated bile acids, by bacterial bile salt hydrolase (BSH), decreases reabsorption efficiency, thus unconjugated bile acids make up 98% of the 5% of bile acids that are excreted daily^(54; 55). This modification is of significant interest, as the production of more readily excreted bile acids, may lead to the increased conversion of cholesterol to bile acids to replace those lost, in turn lowering serum cholesterol⁽⁵⁶⁾.

Cholesterol and Health-span

Intrinsic Ageing

The ageing process has been associated with an increase in both total cholesterol (TC) and LDL-C. For instance, Ericsson et al. (1990) reported an increase in TC from 4.8mmol/L in the young (aged 20-39 years), to 5.14mmol/L in the middle aged (aged 40-59 years), and to 5.44mmol/L in old aged (aged 60-80 years) healthy Scandinavian volunteers⁽⁵⁷⁾. Furthermore, LDL-C increased with age, from 3.37 in the young, to 3.76 in the middle aged, and to 4.05mmol/L in the old aged. Additionally, VLDL-C has been observed to either remain steady or increase with age, while HDL-C appears to be unaffected by the ageing process^(57; 58). Abbott et al. (1993) also found gender influences the lipoprotein profile. For example, females exhibited higher levels of LDL-C, especially in those using oestrogen hormones, and increased HDL-C, whereas VLDL-C was greater in males⁽⁵⁸⁾. The age associated dysregulation of cholesterol metabolism, and accumulation of LDL-C, has been associated with alterations to several key mechanisms, including cholesterol absorption, LDL-C clearance, bile acid synthesis and subsequent intestinal bacterial modification (**Figure 5**)⁽⁵⁾.

Intestinal cholesterol absorption varies greatly between individuals; with estimates ranging from 20.0 to 80.1%^(58; 59). Evidence from murine models suggests cholesterol absorption increases significantly with age⁽⁷⁾. Rodent studies have demonstrated this is mediated by a significant increase in NPC1L1 in both the duodenum and jejunum, while ABCG5/G8 expression is suppressed⁽⁶⁰⁾. It has been estimated this increase in cholesterol absorption and concurrent reduction in efflux from enterocytes, confers a 19-40% increase in cholesterol absorption with age⁽⁶⁰⁾. Moreover, it was observed that high levels of oestrogen upregulated NPC1L1 and ABCG5/G8 mRNA expression. Oestrogen and ageing have been reported to enhance cholesterol absorption via the ER α pathway⁽⁶⁰⁾. It is important to note that these findings have not yet been observed in humans⁽⁵⁹⁾.

Bile acid metabolism is also effected by the ageing process, most significantly, there is a reduction in bile acid synthesis. Wang et al. (2002) found ageing resulted in a significantly reduced biliary bile acid output, from 192-211 μ mol/h/kg to 124-157 μ mol/h/kg in mice. Additionally Wang et al. (2002) demonstrated intrinsic ageing resulted in a reduction in bile acid synthesis, with a 33.3-57.1% and 41.7-56.3% decrease in cholesterol 7 α -hydroxylase (CYP7A1) activity in male and female mice respectively, dependent on dietary and genetic factors⁽⁷⁾. Similarly in humans, an inverse correlation between age and CYP7A1 expression

has been described⁽⁶¹⁾. For instance, cholesterol 7 α -hydroxylation rates were reduced by 50% for individuals over 65 years, compared with individuals below 65 years of age in one Italian cohort⁽⁶²⁾. Bertolotti et al. (1993) estimated by linear regression analysis a 60mg/day (~150 μ mol/day) decline every 10 years, in cholesterol undergoing cholesterol 7 α -hydroxylation⁽⁶²⁾, while Einarsson et al. (1985) estimated an 80mg/day (200 μ mol/day) reduction over the same time period⁽⁶³⁾. Bertolotti et al. (2007) propose the age-related reduction in CYP7A1 expression could be related to the concomitant decline in hepatic nuclear factor 4 and co-activator CYP7A1 promoter binding factor/liver receptor homologue-1 (CPF/LRH-1), mediated by the decline in growth hormone (GH) and insulin-like growth factor (IGF)⁽⁶¹⁾.

Once bile acids reach the small intestine, modification by digestive microflora occurs influencing enterohepatic circulation. Many digestive bacteria produce the enzyme BSH which deconjugates bile acids, decreasing reabsorption efficiency, and enhancing excretion. For example, Tanaka et al. (1999) reported 59 and 98% of *Lactobacillus* and *Bifidobacteria* strains, isolated from faeces are BSH positive⁽⁶⁴⁾. With age there are several changes to the gut microflora, including a decline in the number and species diversity of *Lactobacillus* and *Bifidobacterium*^(8; 65). Therefore it is possible that the age-related decline of these bacterial species, reduces bile acid deconjugation, and in turn reduces the conversion of cholesterol to bile acid. This may play a role in the accumulation of cholesterol with age⁽⁵⁾.

It has also been reported, the clearance rate of LDL-C is affected by the ageing process. The apo B-100 containing lipoproteins, LDL-C and VLDL-C are removed from the blood via hepatic LDLr, for elimination, either by direct efflux or conversion to bile acids. Millar et al. (1995) determined the mean LDL apo B-100 residence time was 2.42 days for younger male adults (mean age 31 \pm 6years) and 3.46 days for older male adults (mean age 61 \pm 10years)⁽⁶⁾. With age, a decline in LDLr activity and/or numbers is thought to be responsible for the reduction in LDL clearance rate and increase in residence time. The reduced conversion of cholesterol by CYP7A1 may play a role in reduction of LDLr. Additionally, proprotein convertase subtilisin/kexin type 9 (PCSK9), a proprotein convertase responsible for the degradation of LDLr has been correlated with age⁽⁶⁶⁾. Interestingly, PCSK9 has also been correlated with BMI, TC, LDL-C, and TAG⁽⁶⁷⁾. Furthermore, Millar et al. (1995) observed, that although VLDL apo B-100 residence time was not effected by age, the production rate of VLDL apo B-100 was correlated with age. The age-related increase in body fat and elevated plasma FFAs were attributed to this increase in VLDL apo B-100 production rate⁽⁶⁾.

Cholesterol metabolism and Diet

Herron et al. (2003) examined the effect of ~640mg/day cholesterol feeding on men aged 18-57 years old, and determined that 37.5% of subjects behaved as hyper-responders, with an increase of ≥ 0.06 mmol/L in TC, while 62.5% behaved as hypo-responders, with an increase in TC of < 0.05 mmol/L. Hyper-responders exhibited a significant 23.0, 7.8 and 18.0% increase in LDL-C and HDL-C, and LDL:HDL ratio respectively, whereas changes to LDL-C, HDL-C, TG and LDL:HDL ratio were not significant for hypo-responders. Interestingly, hyper-responders exhibited elevated LCAT and CETP activity, suggesting the upregulation of RCT as a compensatory mechanism, to reduce the risk of atherosclerosis⁽⁶⁸⁾. Quintao et al. (1971) propose tissue pools of cholesterol may rapidly expand in response to cholesterol feeding, even in the absence of aberrations to plasma cholesterol levels. Typically, there are 2 main mechanisms to compensate for an increase in dietary cholesterol; elevated cholesterol excretion, and decreased cholesterol synthesis⁽⁶⁹⁾. It has been suggested a reduction in cholesterol intake should be considered unnecessary for individuals who have already reduced SFA, and increased PUFA: SFA ratio⁽⁷⁰⁾. Edington et al. (1987) determined a 2-fold increase or decrease in dietary cholesterol in participants who also reduced dietary fat with an increased PUFA:SFA ratio had no effect on TC or LDL-C after 8 weeks⁽⁷⁰⁾. This is due to the significant impact SFA had on serum LDL-C, by influencing several regulatory mechanisms. Firstly, it has been observed there is a reduction in LDLr; resulting in reduced LDL-C clearance and increased LDL-C⁽⁷¹⁾. Mustad et al. (1997) demonstrated an 8 week reduction in SFA resulted in a 10.5% increase in LDLr and subsequent 11.8% decrease in LDL-C. It has been estimated for every 1% increase in LDLr, there is a 0.74% reduction in LDL-C⁽⁷¹⁾. Secondly, SFA may influence cholesterol synthesis⁽⁷²⁾. Glatz & Katan (1993) determined a low PUFA:SFA ratio diet resulted in increased cholesterol synthesis, compared to a high PUFA:SFA ratio diet (1.86mmol/day vs. 1.55mmol/day). Additionally, Jones et al. (1994) demonstrated corn oil increased absolute cholesterol synthesis from 13.9mg/kg/day at baseline, to 21.3mg/kg/day⁽⁷³⁾. Thirdly, it has been demonstrated SFA influences the concentration of CETP. For example, Jansen et al. (2000) observed CETP concentrations were significantly elevated by 12 and 11%, in individuals on a high SFA diet, compared with individuals on the NCEP StepI diet and MUFA diet respectively⁽⁷⁴⁾. Additionally, an elevation of SFA from 8.4 to 11% decreased LCAT activity from 56 to 74nmol/ml/h, which may result in decreased RCT and influence CVD risk⁽⁷⁵⁾.

Cholesterol metabolism and Disease

The age related accumulation of both TC and LDL-C has been associated with the pathogenesis of several diseases^(76; 77; 78). In a meta-analysis of 62 studies, a 17.5% reduction in relative risk for all-cause mortality for every 1mmol/L decrease in TC was reported, while each 1mmol/L decrease in LDL-C was associated with a 15.6% relative risk reduction for all-cause mortality⁽⁷⁹⁾.

CHD has been associated with significantly elevated TC, LDL-C, TAG, apo B, and Lp(a) and reduced HDL-C and apo A-I in both American males and females⁽⁷⁸⁾. Gould et al. (2007) reported a 24.5 and 29.5% reduction in relative risk for CHD related mortality and CHD event respectively for every 1mmol/L decline in TC. Additionally, each 1mmol/L decline in LDL-C was associated with a 28.0 and 26.6% decline in relative risk for CHD related mortality and CHD event respectively⁽⁷⁹⁾. Sharrett et al. (2001) found each 1mmol/L increase in LDL-C was associated with a relative risk for CHD of between 1.36-1.44 for males and 1.19-1.32 for females, while a 0.4mmol/L increase in HDL-C was associated with a 0.64-0.72 and 0.64-0.76 relative risk for CHD for males and females, respectively⁽⁷⁸⁾. Interestingly, the decreased risk for mortality with reduced TC has been observed to decline with age. For example, a meta-analysis conducted by Lewington et al. (2007) concluded a 1mmol/L reduction in TC was associated with an ~50% reduction in IHD mortality for 40-49 year olds, which decreased to an ~33% reduction for 50-69 year olds and ~17% for 70-89 year olds⁽⁸⁰⁾.

Intriguingly, several studies have described an inverse relationship between cholesterol and mortality⁽⁸¹⁾. Lv et al. (2015) observed a 19% decrease in relative risk for each 1mmol/L increase in LDL-C in Chinese ≥ 80 year olds. Additionally, a 40% lower risk for mortality was associated with those with abnormally high LDL-C (≥ 3.37 mmol/L), compared to those with a lower plasma concentration of LDL-C⁽⁸²⁾. Similarly, Takata et al. (2014) observed a 0.8% decrease in mortality for each 1mg/dL (0.026mmol/L) increase in LDL-C in Japanese ≥ 80 year olds. In addition, each 1mg/dL increase in TC was associated with a 0.9% reduction in mortality⁽⁸³⁾.

Targeting Cholesterol Metabolism as a Therapeutic Strategy

There are many strategies utilised in the treatment of hypercholesterolaemia. These can be used singularly or in combination, and include changes to diet and exercise regimes, pharmacological intervention, and dietary supplementation^(9; 11; 84; 85).

Dietary Intervention

A reduced intake of SFA, and increase in MUFA and PUFA may be useful in maintaining a healthy lipid profile, and reducing CHD risk⁽¹¹⁾. An 8 week reduction in SFA was observed to reduce TC and LDL-C by 9.3 and 11.8% respectively in healthy males and females aged between 20 and 65 years old⁽⁷¹⁾. Additionally, Berry et al. (1991) reported a 12 week MUFA or PUFA diet reduced TC by ~10 and ~16% respectively, while reducing LDL-C by 14 and 21%⁽⁸⁶⁾. A meta-analysis of 13,614 participants participating in diets where SFA was replaced with PUFA, found that on average there was a 0.76mmol/L (29mg/dL) reduction in TC, and that each 1mmol/L reduction in TC was associated 24% reduced risk for a CHD event. Moreover, each additional year of diet was related to a further 9.2% risk reduction⁽¹¹⁾.

Additionally, the use of phytosterols is recommended as a therapeutic strategy for the treatment of hypercholesterolaemia. It is estimated a 2g/day dose of phytosterols lowers LDL-C by 10%⁽⁸⁷⁾. A meta-analysis of 124 studies concluded, 0.6-3.3g/day of phytosterols reduced LDL-C by 6-12%, with stanols and sterols showing similar LDL-C lowering efficacies⁽⁸⁵⁾. The effect of phytosterols has been observed to taper off at 2-3g/day⁽⁸⁷⁾, therefore 2mg/day has been recommended to aid in the treatment of hypercholesterolaemia⁽⁸⁸⁾.

Pharmacological Intervention

Lipid lowering drugs are often utilised in the treatment of CVD (**Figure 6**). Statins are often recommended for those diagnosed with CVD, or those with a high risk of developing the disease, due to their tolerability and efficacy⁽⁸⁹⁾. Statins act by competitively inhibiting HMG CoA reductase, the rate limiting enzyme of cholesterol synthesis in order to lower serum cholesterol, thus reducing CVD risk⁽⁹⁰⁾. Simvastatin reduced LDL-C by 33% after 12 weeks, in patients with a history of myocardial infarction, while atorvastatin exhibited a 49% reduction⁽⁹¹⁾. A meta-analysis of 4 trials demonstrated standard-dose statin therapy reduced LDL-C by 22%, to a mean of 2.59mmol/L (101mg/dL), while high-dose statin therapy reduced LDL-C by 42%, to a mean of 1.92mmol/L (75mg/dL)⁽⁹²⁾. The standard dose therapy was associated with a coronary death or myocardial event rate of 9.4%, whereas high dose therapy was associated with an 8.0% event rate. Significantly, overall statin therapy resulted in a 16% odds reduction in coronary death or myocardial infarction⁽⁹²⁾. Ridker et al. (2005) observed that patients whose statin therapy reduced LDL-C to <70mg/dL (<1.81mmol/dL), had a reduced rate of recurrent myocardial infarction or death from coronary events (2.7

events per 100 person-years), than individuals whose statin therapy did not reach this goal (4.0 events per 100 person-years)⁽⁹³⁾. However, it has also been demonstrated non-statin therapy (diet, bile acid sequestrants and ileal bypass surgery) reduced CHD risk equally to that of statins, with each 1% reduction in LDL-C corresponding to a 1% decrease in CHD risk⁽¹⁰⁾. Statins can also be used in combination with drugs such as Ezetimibe, which targets NPC1L1 to lower cholesterol absorption. Cannon et al. (2015) observed that combination therapy lowered LDL-C by a further 24% than statin therapy alone. Combination therapy also significantly reduced myocardial infarction and ischemic stroke risk⁽⁸⁴⁾.

Targeting the Gut Microbiome

There is emerging evidence indicating the important role for the gut microbiome in regulating many biological systems, including cholesterol metabolism^(94; 95). Ageing is associated with the alteration of the gut microbiome. These changes differ between individuals and populations, as diet, lifestyle, host health and antibiotic use likely play a role in composition^(96; 97). However, it has been observed *Lactobacillus* and *Bifidobacterium* species diversity and total count decline with age^(8; 65). Supplementation with probiotics may have the ability to partially ameliorate changes associated with ageing, such as immunosenescence, neurodegeneration, carcinogenic transformation, and hypercholesterolaemia^(98; 99).

With regards to hypercholesterolaemia treatment, supplementation with probiotics has been associated with a decline in TC and LDL-C⁽⁹⁾. A meta-analysis of probiotic administration studies concluded probiotic administration resulted in a mean 6.40mg/dL (0.17mmol/L) reduction in TC, and 4.90mg/dL (0.13mmol/L) reduction in LDL-C. Additionally probiotics were observed to have beneficial effects on TC and LDL-C for individuals with high, borderline high, and normal cholesterol levels⁽⁹⁾. Probiotics have also been an effective treatment strategy for individuals with underlying conditions. For example, Bernini et al. (2016) observed ingestion of 80ml probiotic milk containing 3.4×10^8 colony forming units (CFU)/ml of *Bifidobacterium lactis* HN019 for 45 days, significantly lowered TC from 209 to 194mg/dL (5.40mmol/L and 5.02mmol/L), and LDL from 128.5 to 111mg/dL (3.32mmol/L to 2.87mmol/L) in patients with metabolic syndrome, in addition to lowering BMI. Moreover, the proinflammatory cytokines TNF- α and IL-6 were significantly reduced with probiotic treatment⁽¹⁰⁰⁾.

There are several proposed mechanisms to explain the lipid-lowering ability of probiotics⁽¹⁰¹⁾. For example, some probiotics produce the bile acid deconjugating enzyme BSH, which may

increase bile acid excretion, and upregulate the conversion of cholesterol to bile acids to replace those lost⁽¹⁰²⁾. It has also been observed that probiotics increase short chain fatty acids (SCFA), which inhibit the hepatic rate limiting enzyme in cholesterol synthesis, HMG CoA reductase⁽¹⁰¹⁾. Furthermore, there may be a reduction in cholesterol absorption due to the assimilation and possible incorporation of cholesterol into the bacterial cell membrane⁽¹⁰³⁾. Additionally, some bacteria may act by destabilising cholesterol micelles resulting in the coprecipitation of cholesterol with deconjugated bile acids⁽¹⁰³⁾.

Applying Systems Biology to our Understanding of Cholesterol Metabolism and Ageing

It is apparent from our discussion of cholesterol metabolism, that it is an exceptionally complex system with a multitude of interacting mechanisms^(104; 105). Many of these mechanisms interact via positive and negative feedback regulators, the dynamics of which is not trivial. Our understanding of this complexity is further compounded by ageing which alters the behaviour of these regulatory mechanisms, thus modifying the overall dynamics of whole-body cholesterol metabolism. Moreover, changes to extrinsic factors such as diet exert a significant influence on the overall behaviour of this system. Thus, it is imperative if we are to gain a more in-depth appreciation of cholesterol metabolism and its interaction with ageing, that we investigate this system in a more integrated manner. The systems biology paradigm contrasts with the more traditional reductionist approach, commonly used in nutrition research, and offers a more integrated way to study this multifaceted system^(14; 106; 107). A fundamental element of this paradigm shift is the close coupling of computational modelling with experimental work^(108; 109; 110; 111). Our group has used computational systems biology to investigate cholesterol metabolism and ageing⁽¹¹²⁾. Firstly, we constructed a whole-body model of cholesterol metabolism^(113; 114). The model is defined by the key components that characterise whole body cholesterol balance, namely cholesterol ingestion, excretion and synthesis together with LDLr receptor turnover and RCT. Using *in silico* experiments we explored the impact of ageing on these fundamental elements of cholesterol metabolism. Firstly, we investigated the influence of ageing on cholesterol absorption and it was found that for every 10% increment in the rate of cholesterol absorption, there was a concomitant increase of 12.5 mg/dL in LDL-C. The model also revealed that increasing cholesterol absorption from 50% to 80% by age 65 years, resulted in a 34 mg/dL increase in plasma LDL-C. In addition, the model found that reducing the number of hepatic LDLr had a

significant impact on the system. An increase of 116 mg/dL in LDL-C was observed by 65 years in response to a reduction of 50% in their numbers. Thus, emphasizing LDLr maintenance as a key component in maintaining cholesterol metabolism during ageing. This model is coded in the systems biology markup language (SBML) and archived in the BioModels database a repository for models encased in SBML exchange framework (<http://www.ebi.ac.uk/biomodels-main/BIOMD0000000434>). This means the model is straightforward to update and adapt, a feature exploited by other groups working in this area⁽¹¹⁵⁾. We recently implemented significant updates to the model described above for the following reasons⁽¹¹⁶⁾. The model was lacking several key mechanisms which recent experimental evidence has stressed are central to the regulation of cholesterol metabolism. Briefly, these processes included plasma membrane receptors, and *in vivo* and intestinal microfloral enzymes. Therefore, it was deemed cogent that these 96 additional mechanisms were incorporated into the original model⁽⁵⁾. We found that our updated model behaved as a hypo-responder to excessive cholesterol feeding. Moreover, the model was utilised to investigate the effects of ageing coupled with three different CETP genotypes. Ageing in the presence of a genotype conferring low CETP activity resulted in a 0.6% decrease in LDL-C after 1000 hours. In comparison, ageing with a genotype indicative of high CETP activity, provoked a 1.6% increase in LDL-C levels (**Figure 7**). Thus, our new model consolidates experimental findings, which emphasize the significance of CETP genotypes in health-span⁽¹¹⁶⁾.

Complimenting Computational models with Simulated Digestive tracts

In order to complement our *in silico* studies of cholesterol metabolism, we aim to develop a simulator of the human digestive tract. The digestive track simulator will be used to refine and inform our computational model. One example is the simulator of the human intestinal microbial ecosystem (SHIME) reactor, which represents the length of the human digestive system, with several closed reactors used to represent the differing conditions (microbial ecosystem, pH, enzymes, etc.) of each section of the digestive tract⁽¹¹⁷⁾. The flexible nature of the SHIME reactor means that can be easily adapted. This may include additional reactors to more accurately represent the human digestive tract^(118; 119). Simulated digestive tracts, such as the SHIME reactor provide a suitable test environment to supplement experimental data on potential therapeutic strategies. For example, De Smet et al. (1998) determined administration of *Lactobacillus* to pigs for 4 weeks decreased TC and LDL-C by 15 and 24%, respectively.

Additionally, TC and LDL-C reduced a further 18 and 34% 3 weeks after treatment⁽¹²⁰⁾. Follow-up experimentation using the SHIME reactor was able to provide details of the intestinal bacterial population and overall metabolic activity associated with this treatment⁽¹¹⁸⁾. Recently, a continuous gut adhesion model was developed using mucin-alginate beads to immobilise bacteria, which may more effectively represent the gut lining and intestinal lumen. This system was also used to examine bacterial colonisation following probiotic treatment⁽¹²¹⁾. The TNO gastro-intestinal model (TIM) can be used to examine compound absorption⁽¹²²⁾. This model simulates ingestion of food and water, digestive enzyme and bile salt release, peristalsis, body temperature, and transit through the stomach and small intestine (TIM-1) and large intestine (TIM-2) under computer control. The model can be altered to represent a different species, such as a human or pig, or can represent varying age, such as infant, adult and older people⁽¹²²⁾. This is particularly significant for the augmentation and refinement of our computational model. It is hoped that utilizing both these approaches in tandem will help to significantly deepen our understanding of cholesterol metabolism and its relationship with ageing.

Discussion

The dysregulation of cholesterol metabolism accompanies the ageing process; this may play a role in disease pathogenesis. We have outlined how the age related dysregulation of cholesterol metabolism is influenced by many factors. Specifically we discussed i) an increase in LDL-C residence time due to reduction in LDLr and concomitant decline in LDL-C clearance, ii) a decline in CYP7AI activity which reduces bile acid synthesis, one pathway for cholesterol removal, iii) a reduction in digestive bacteria which modify bile acids that reduce reabsorption and promotes excretion and iv) a potential increase in cholesterol absorption due to an increase in NPC1L1 and decrease in ABCG5/G8, as observed in murine models. Cholesterol metabolism is also influenced by diet. For example, a meta-analysis of 67 trials revealed that soluble fibre could lower TC and LDL-C by 0.045 and 0.057mmol/L/g soluble fibre respectively⁽¹²³⁾. There are a variety of mechanisms that dietary fibre could influence; for example, bile acid micelle formulation, fat excretion, intestinal motility, SCFA production, and absorption of macronutrients^(124; 125; 126). Interestingly, diet may not only impact cholesterol metabolism directly, and may also interact with the gut microbiome⁽¹²⁷⁾. David et al. (2014) describe that short term changes to diet could significantly influence both

the microbial community structure and bacterial gene expression. For example, a 5 day animal based-diet significantly altered the abundance of 22 clusters, elevated faecal bile acid concentration, and increased BSH expression⁽¹²⁸⁾. Evard et al. (2014) describe that high fat feeding significantly affected 20 genera. This may have been influenced by a decline in the antimicrobial peptide regenerating islet-derived protein 3 gamma (Reg3g). Interestingly, prebiotic treatment increased Reg3g ~6-fold within the colon. Targeting the digestive microbiota with pre- and probiotics is a potential avenue for the treatment of diseases including hypercholesterolaemia. For instance, meta-analysis of 11 clinical trials demonstrated that probiotic intervention was associated with an average reduction in TC and LDL-C of 0.17mmol/L and 0.22mmol/L respectively. Studying these mechanisms underpinning these complex interactions of intrinsic ageing and diet on cholesterol metabolism, can be challenging. In recent years, nutrition research has benefitted from using computational modelling to study the complexities associated with ageing and cholesterol metabolism. This approach provides a meaningful framework for handling this complexity. Modelling enables hypothesis exploration, and allows us to predict the behaviour of cholesterol metabolism over time. We used computational modelling to construct a whole body mathematical model of cholesterol metabolism and its age associated dysregulation. Encased within this template we incorporated many key mechanisms, including LDLr turnover, intestinal cholesterol absorption, and endogenous cholesterol synthesis. Using our *in silico* 20 year old, a number of mechanisms were explored. By increasing cholesterol absorption from 50% to 80% by 65 years, we were able to show that LDL-C increased by 34 mg/dL from its baseline value of 100mg/dL at 20 years of age in a healthy adult male. However, the key finding of the model centred on hepatic LDLr. Using the model we were able to show that by decreasing the activity of the LDLr to 50% by age 65 years, this produced a rise in LDL-C of 116mg/dL, from a base line value of 100 mg/dL at age 20 years in a healthy male. More recently, we updated the model to include a further 96 mechanisms. The updated model behaved as a hypo-responder to increased dietary cholesterol, whereas SFA feeding elevated LDL-C, in addition to hepatic and peripheral pools of esterified cholesterol. Furthermore, the age related increase in LDL-C was reduced in the presence of low CETP activity. In addition to exploring these aspects of cholesterol metabolism it is possible that computational modelling could be used to examine the role of the gut microbiome on host health⁽¹²⁹⁾. Moreover computational models are an effective way of complimenting simulated digestive tracts, such as the SHIME reactor which is a useful tool for investigating the microbial community and metabolic activity of each section of the

digestive tract in response to ingested substrates, something which is difficult to achieve *in vivo*⁽¹³⁰⁾. This is particularly significant as recent evidence has indicated that the gut microbiome could have a significant impact health-span and may provide a potential therapeutic target to treat age associated hypercholesterolaemia.

Figures

Figure 1 UK life expectancy by year of birth. Data from Clio-Infra⁽¹³¹⁾.

Figure 2 UK population by age and gender in 1982 and 2012. Data from United Nations Statistics Division⁽¹³²⁾.

Figure 3 UK causes of death by age. Data from British Heart Foundation⁽¹³³⁾.

Figure 4 Modelling Overview. (1) Identify the system to model and hypothesis formation. (2) Identify pre-existing models; using the BioModels Database, a repository for peer reviewed models. (3) If no model of the system of interest exists: produce a network diagram. If a model does exist: download model and move to step 5, then step 7. (4) Establish mathematical framework. (5) Identify a suitable modelling tool; there are several available including: COPASI, which we utilized in our updated model of cholesterol metabolism⁽¹¹⁶⁾, CellDesigner, Mathematica and MATLAB. (6) Obtain initial concentrations of species, rate laws and kinetic data to construct the model. The online resources BRENDA and SABIO-RK provide a substantial volume of kinetic data. (7) Run simulations. (8) Validate the model. (9) Explore the hypotheses, and determine if the model accurately represents the biological system, and can be used to make predictions, or if the model needs refining. (10) Conduct further wet laboratory experiments based upon model output. (11) Code the model in the exchange format, Systems Biology Markup Language (SBML) and deposit in the BioModels Database. Adapted from Mc Auley and Mooney (2015)⁽¹³⁴⁾.

Figure 5 Overview of age-related changes to cholesterol metabolism.

Figure 6 UK prescription drugs for cardiovascular disease. Data from British Heart Foundation⁽¹³³⁾.

Figure 7 Model simulation of ageing in the presence of CETP genotypes. Taken from Morgan et al. (2016)⁽¹¹⁶⁾.

Acknowledgements

A. E. M. was funded by a University of Chester PhD scholarship.

Financial Support

This research received no specific grant from any funding agency in the public, commercial or not-for-profit sectors.

Conflict of Interest

None.

Authorship

A.E.M. and M.T.MA. drafted the manuscript. K.M.M. and N.A.P. reviewed and revised the manuscript.

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